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(54) Title: NOVEL MAMMALIAN CALCIUM CHANNELS AND RELATED PROBES, CELL LINES AND METHODS

(57) Abstract: Sequences and partial sequences for three types of mammalian (human and rat sequences identified) T-type calcium channel subunits which we have labeled as the  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits are provided. Knowledge of the sequence of these calcium channels permits the localization and recovery of the complete sequence from human cells, and the development of cell lines which express the novel calcium channels of the invention. These cells may be used for identifying compounds capable of acting as agonists or antagonists to the calcium channels.

## NOVEL MAMMALIAN CALCIUM CHANNELS AND RELATED PROBES, CELL LINES AND METHODS

### TECHNICAL FIELD

The invention relates to T-type calcium channel encoding sequences,  
5 expression of these sequences, and methods to screen for compounds which  
antagonize calcium channel activity. The invention is also related to molecular tools  
derived from knowledge of the molecular structure of T-type calcium channels.

### BACKGROUND OF THE INVENTION

The rapid entry of calcium into cells is mediated by a class of proteins called  
10 voltage-gated calcium channels. Calcium channels are a heterogeneous class of  
molecules that respond to depolarization by opening a calcium-selective pore through  
the plasma membrane. The entry of calcium into cells mediates a wide variety of  
cellular and physiological responses including excitation-contraction coupling,  
hormone secretion and gene expression. In neurons, calcium entry directly affects  
15 membrane potential and contributes to electrical properties such as excitability,  
repetitive firing patterns and pacemaker activity. Miller, R.J. (1987) "Multiple  
calcium channels and neuronal function." *Science* 235:46-52. Calcium entry further  
affects neuronal functions by directly regulating calcium-dependent ion channels and  
modulating the activity of calcium-dependent enzymes such as protein kinase C and  
20 calmodulin-dependent protein kinase II. An increase in calcium concentration at the  
presynaptic nerve terminal triggers the release of neurotransmitter. Calcium entry  
also plays a role in neurite outgrowth and growth cone migration in developing  
neurons and has been implicated in long-term changes in neuronal activity.

In addition to the variety of normal physiological functions mediated by  
25 calcium channels, they are also implicated in a number of human disorders. Recently,  
mutations identified in human and mouse calcium channel genes have been found to  
account for several disorders including, familial hemiplegic migraine, episodic ataxia  
type 2, cerebellar ataxia, absence epilepsy and seizures. Fletcher, *et al.* (1996)  
"Absence epilepsy in tottering mutant mice is associated with calcium channel  
30 defects." *Cell* 87:607-617; Burgess, *et al.* (1997) "Mutation of the Ca<sup>2+</sup> channel

$\beta$  subunit gene *Cchb4* is associated with ataxia and seizures in the lethargic (lh) mouse." *Cell* 88:385-392; Ophoff, *et al.* (1996) "Familial hemiplegic migraine and episodic ataxia type-2 are caused by mutations in the  $\text{Ca}^{2+}$  channel gene *CACNL1A4*." *Cell* 87:543-552; Zhuchenko, O. *et al.* (1997) "Autosomal dominant cerebellar ataxia (SCA6) associated with the small polyglutamine expansions in the  $\alpha_{1A}$ -voltage-dependent calcium channel." *Nature Genetics* 15:62-69.

The clinical treatment of some disorders has been aided by the development of therapeutic calcium channel antagonists. Janis, *et al.* (1991) in *Calcium Channels: Their Properties, Functions, Regulation and Clinical Relevance*. CRC Press, London.

Native calcium channels have been classified by their electrophysiological and pharmacological properties as T, L, N, P and Q types (for reviews see McCleskey, *et al.* (1991) "Functional properties of voltage-dependent calcium channels." *Curr. Topics Membr.* 39: 295-326, and Dunlap, *et al.* (1995) "Exocytotic  $\text{Ca}^{2+}$  channels in mammalian central neurons." *Trends Neurosci.* 18:89-98.). T-type (or low voltage-activated) channels describe a broad class of molecules that activate at negative potentials and are highly sensitive to changes in resting potential. The L, N, P and Q-type channels activate at more positive potentials and display diverse kinetics and voltage-dependent properties. There is some overlap in biophysical properties of the high voltage-activated channels, consequently pharmacological profiles are useful to further distinguish them. L-type channels are sensitive to dihydropyridine (DHP) agonists and antagonists, N-type channels are blocked by the *Conus geographus* peptide toxin,  $\omega$ -conotoxin GVIA, and P-type channels are blocked by the peptide  $\omega$ -agatoxin IVA from the venom of the funnel web spider, *Agelenopsis aperta*. A fourth type of high voltage-activated Ca channel (Q-type) has been described, although whether the Q- and P-type channels are distinct molecular entities is controversial (Sather *et al.* (1993) "Distinctive biophysical and pharmacological properties of class A (B1) calcium channel  $\alpha_1$  subunits." *Neuron* 11:291-303; Stea, *et al.* (1994) "Localization and functional properties of a rat brain  $\alpha_{1A}$  calcium channel reflect similarities to neuronal Q- and P-type channels." *Proc Natl Acad Sci (USA)* 91:10576-10580; Bourinet, E. *et al.* (1999) *Nature Neuroscience* 2:407-415). Several types of calcium conductances do not fall neatly into any of the above categories and there is variability of properties even within a category suggesting that additional calcium channels subtypes remain to be classified.

Biochemical analyses show that neuronal high-threshold calcium channels are heterooligomeric complexes consisting of three distinct subunits ( $\alpha_1$ ,  $\alpha_2\delta$  and  $\beta$ ) (reviewed by De Waard, *et al.* (1997) in *Ion Channels*, Volume 4, edited by Narahashi, T. Plenum Press, New York). The  $\alpha_1$  subunit is the major pore-forming subunit and contains the voltage sensor and binding sites for calcium channel antagonists. The mainly extracellular  $\alpha_2$  subunit is disulphide-linked to the transmembrane  $\delta$  subunit and both are derived from the same gene and are proteolytically cleaved *in vivo*. The  $\beta$  subunit is a non-glycosylated, hydrophilic protein with a high affinity of binding to a cytoplasmic region of the  $\alpha_1$  subunit. A fourth subunit,  $\gamma$  is unique to L-type Ca channels expressed in skeletal muscle T-tubules. The isolation and characterization of  $\gamma$ -subunit-encoding cDNAs is described in U.S. Patent No. 5,386,025 which is incorporated herein by reference.

Molecular cloning has revealed the cDNA and corresponding amino acid sequences of six different types of  $\alpha_1$  subunits ( $\alpha_{1A}$ ,  $\alpha_{1B}$ ,  $\alpha_{1C}$ ,  $\alpha_{1D}$ ,  $\alpha_{1E}$  and  $\alpha_{1S}$ ) and four types of  $\beta$  subunits ( $\beta_1$ ,  $\beta_2$ ,  $\beta_3$  and  $\beta_4$ ) (reviewed in Stea, A., Soong, T.W. and Snutch, T.P. (1994) "Voltage-gated calcium channels." in *Handbook of Receptors and Channels*. Edited by R.A. North, CRC Press). A comparison of the amino acid sequences of these  $\alpha_1$  subunits is included in this publication, which is incorporated herein by reference. PCT Patent Publication WO 95/04144, which is incorporated herein by reference, discloses the sequence and expression of  $\alpha_{1E}$  calcium channel subunits.

As described in Stea, A. *et al.* (1994) (*supra*), the  $\alpha_1$  subunits are generally of the order of 2000 amino acids in length, ranging from 1873 amino acids in  $\alpha_{1S}$  derived from rabbit to 2424 amino acids in  $\alpha_{1A}$  derived from rabbit. Generally, these subunits contain 4 internal homologous repeats (I-IV) each having six putative alpha helical membrane spanning segments (S1-S6) with one segment (S4) having positively charged residues every 3rd or 4th amino acid. There are a minority of a splice variant exceptions. Between domains II and III there is a cytoplasmic domain which is believed to mediate excitation-contraction coupling in  $\alpha_{1S}$  and which ranges from 100-400 amino acid residues among the subtypes. The domains I-IV make up roughly 2/3 of the molecule and the carboxy terminus adjacent to the S6 region of domain IV is believed to be on the intracellular side of the calcium channel. There is a consensus motif (QQ-E-L-GY-WI-E) in all of the subunits cloned and described in Stea, A. *et al.*



(supra) downstream from the domain I S6 transmembrane segment that is a binding site for the B subunit.

PCT publication WO 98/38301, which describes the work of the inventors herein, and which is incorporated herein by reference, reports the first description of the molecular composition of T-type calcium channel  $\alpha_1$  subunits. The present application describes full-length genes for 3 mammalian subtypes,  $\alpha_{1G}$ ,  $\alpha_{1H}$ , and  $\alpha_{1I}$  associated with T-type calcium channels.

In some expression systems the high threshold  $\alpha_1$  subunits alone can form functional calcium channels although their electrophysiological and pharmacological properties can be differentially modulated by coexpression with any of the four  $\beta$  subunits. Until recently, the reported modulatory affects of  $\beta$  subunit coexpression were to mainly alter kinetic and voltage- dependent properties. More recently it has been shown that  $\beta$  subunits also play crucial roles in modulating channel activity by protein kinase A, protein kinase C and direct G-protein interaction. (Bourinet, *et al.* (1994) "Voltage-dependent facilitation of a neuronal  $\alpha_1C$  L-type calcium channel." *EMBO J.* 13: 5032-5039; Stea, *et al.* (1995) "Determinants of PKC- dependent modulation of a family of neuronal calcium channels." *Neuron* 15:929-940; Bourinet, *et al.* (1996) "Determinants of the G-protein-dependent opioid modulation of neuronal calcium channels." *Proc. Natl. Acad. Sci. (USA)* 93: 1486-1491.)

Because of the importance of calcium channels in cellular metabolism and human disease, it would be desirable to identify the remaining classes of  $\alpha_1$  subunits, and to develop expression systems for these subunits which would permit the study and characterization of these calcium channels, including the study of pharmacological modulators of calcium channel function.

## DISCLOSURE OF THE INVENTION

The present invention provides sequences for a novel mammalian calcium channel subunits of T-type calcium channels, which we have labeled as  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits. Knowledge of the sequences of these calcium channel subunits may be used in the development of probes for mapping the distribution and expression of the subunits in target tissues. In addition, as the molecular structure of the  $\alpha_1$  subunits of these T-type calcium channels has been elucidated, it is possible to identify those

portions which reside extracellularly and thus to design peptides to elicit antibodies which can be employed to assess the location and level of expression of T-type calcium channels. In addition, these subunits, either alone or assembled with other proteins, can produce functional calcium channels, which can be evaluated in model cell lines to determine the properties of the channels containing the subunits of the invention. These cell lines can be used to evaluate the effects of pharmaceuticals and/or toxic substances on calcium channels incorporating  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits. The resulting identified compounds are useful in treating conditions where undesirable T-type calcium channel activity is present. These conditions include epilepsy, sleep disorders, mood disorders, cardiac hypertrophy and arrhythmia and hypertension, among others. In addition, antisense and triplex nucleotide sequences can be designed to inhibit the production of T-type calcium channels.

In some embodiments of the methods and products of this invention, the  $\alpha_1$  subunits are other than those encoded by SEQ ID NO: 17; or, alternatively, are other than those encoded by SEQ ID NO: 17 and by the full length sequences of which SEQ ID NO: 19 and 21 are part. Other embodiments of the methods and products of this invention exclude probes representing portions of or all of SEQ ID NO: 13-21; or, alternatively, exclude probes representing portions of or all of SEQ ID NO: 1-22.

## BRIEF DESCRIPTION OF THE DRAWINGS

Figs. 1A and B show a comparison of the waveforms and current voltage relationship for  $\alpha_{1G}$ ;

Figs. 2A and B show a comparison of the waveforms and current voltage relationship for  $\alpha_{1I}$  calcium channels.

Fig. 3 shows a comparison of the steady state inactivation profiles of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels.

Figs. 4A-C show a comparison of the inactivation kinetics of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels.

Figures 5A and 5B show the construction of the human  $\alpha_{1G}$  cDNA complete sequence from partial clones.

Figure 6 shows the nucleotide and deduced amino acid sequence of human T-type calcium channel  $\alpha_{1G}$ .

Figure 7 shows a comparison of the waveforms and current voltage relationship for human  $\alpha_{1G}$  calcium channel.

Figure 8 shows the characteristic pore pattern for T-type channels.

## 5 MODES OF CARRYING OUT THE INVENTION

The present invention includes the following aspects for which protection is sought:

- (a) novel mammalian (including human) calcium channel subunits and
- 10 DNA sequences encoding such subunits. Specifically, the invention encompasses an at least partially purified DNA molecule comprising a sequence of nucleotides that encodes an  $\alpha_1$  subunit of a T-type calcium channel, and such  $\alpha_1$  subunits *per se*. It will be appreciated that polymorphic variations may be made or may exist in the DNA of some individuals leading to minor deviations in the DNA or amino acids sequences
- 15 from those shown which do not lead to any substantial alteration in the function of the calcium channel. Such variations, including variations which lead to substitutions of amino acids having similar properties are considered to be within the scope of the present invention. Thus, in one embodiment, the present application claims DNA molecules which encode  $\alpha_1$  subunits of mammalian T-type calcium channels, and
- 20 which hybridize under conditions of medium (or higher) hybridization stringency with one or another of the specific sequences disclosed in this application. This level of hybridization stringency is generally sufficient given the length of the sequences involved to permit recovery of the subunits within the scope of the invention from mammalian DNA libraries.

- 25 Alternatively, the T-type calcium channels of the invention are recognized by their functional characteristic of low voltage gating along with defined structural characteristics which classify them as  $\alpha_1$  calcium channel subunits and also characterize them as of the T-type. By virtue of the present invention, these characteristics have been elucidated as follows:

- 30 One distinguishing feature of the  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  T-type channels over other types of calcium channels and sodium channels is that the pore region (P-region) in each of the four structural domains contains a diagnostic amino acid sequence implicated in channel permeability. Figure 8 shows that the T-type channels contain the residues glutamate/glutamate/aspartate/aspartate (single letter amino acid code:

EEDD) in their P-regions (in domains I-IV). In contrast, figure 8 shows that in sodium (Na) channels the P-region of the four domains contains the residues: aspartate/glutamate/lysine/alanine (single letter amino acid code: DEKA), while high threshold calcium channels such as the L-type channel contain the residues:  
5 glutamate/glutamate/glutamate/glutamate (single letter amino acid code: EEEE). The  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  T-type channels are also distinct in this region compared to other types of ion channels including the *C. elegans* C11D2.6 and C27F2.3 and the rat NIC-channel (Figure 8).

A second distinguishing characteristic of the  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  T-type channels  
10 compared to other types of calcium channels is that they do not contain a  $\beta$  subunit binding consensus sequence in the cytoplasmic linker separating domains I and II. In contrast, all high threshold calcium channels contain a consensus sequence (single letter amino acid code: QQ-E-L-GY-WI-E) shown to physically interact with the calcium channel  $\beta$  subunit (Pragnell, M., De Waard, M., Mori, Y., Tanabe, T., Snutch,  
15 T.P. & Campbell, K.P., 1994, Nature 368:67-70). Thus, it appears the presence of a  $\beta$  subunit does not modify activity, nor is its presence required.

A third distinguishing characteristic of the ( $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  T-type channels is that they do not possess an EF-hand calcium binding motif in the region carboxyl to domain IV S6. In contrast, all high threshold calcium channels contain a consensus  
20 sequence that is closely related to the EF-hand domain found in certain calcium binding proteins (de Leon, M., Wang, Y., Jones, L., Perez-Reyes, E., Wei, X., Soong, T.W., Snutch, T.P. & Yue, D.T., 1995, Science 270: 1502-1506).

Thus, as defined herein, "T-type calcium channel  $\alpha_1$  subunits" refers to subunits which contain these structural characteristics.

25 Alternatively, the T-type  $\alpha_1$  subunit molecules can be defined by homology to the human and rat nucleotide and amino acid sequences described herein. Thus, T-type  $\alpha_1$  subunits will typically have at least 50% and preferably 70% homology in terms of amino acid sequence or encoding nucleotide sequence to the sequences set forth in SEQ ID NOS. 23-28 herein or those shown in Figure 6. Preferably, the  
30 homology will be at least 80%, more preferably 90%, and most preferably 95%, 97%, 98% or 99%.

Relative homology may also be defined in terms of specific regions; as set forth above, certain regions of T-type channel  $\alpha_1$  subunits have very high homologies while other regions, such as the cytoplasmic region between domains II and III have less homology. Thus, T-type  $\alpha_1$  subunits will have over 75% homology, preferably over 85% or over 95% homology, more preferably over 98% homology in domains I-IV to those of SEQ ID NO: 23-28 or Figure 6. The degree of homology in the cytoplasmic region between domains II and III may be substantially less, *e.g.*, only 25% homology, preferably 50% homology or more preferably 60% homology. Similarly, the intracellular region downstream of domain IV may be less homologous than those within domains I-IV.

(b) polynucleotide sequences useful as probes in screening human cDNA libraries for genes encoding these novel calcium channel subunits. These probes can also be used in histological assay to determine the tissue distribution of the novel calcium channel subunits.

As set forth above, the elucidation herein of the structural features of T-type subunits permits the selection of appropriate probes by selecting portions of the encoding nucleotide sequence that are particularly characteristic of this type. As set forth above, for example, T-type subunits have particular patterns of amino acids in the pore forming units as set forth in Figure 8. Alternatively, multiple probes might be used to distinguish other subunits, such as probes which represent the  $\beta$ -binding domain missing from the T-type  $\alpha_1$  subunits combined with a probe representing a consensus sequence for calcium channel  $\alpha$  subunits in general.

(c) at least partially purified  $\alpha_1$  subunits and related peptides for mammalian T-type calcium channels. These proteins and peptides can be used to generate polyclonal or monoclonal antibodies to determine the cellular and subcellular distribution of T-type calcium channel subunits.

Again, by virtue of the elucidation of the amino acid sequence of T-type  $\alpha_1$  subunits, it is well within the ordinary skill in the art to determine which regions of the channel are displayed extracellularly and to select these regions for the generation of antibodies.

(d) eukaryotic cell lines expressing the novel calcium channel subunits. These cell lines can be used to evaluate compounds as pharmacological modifiers of the function of the novel calcium channel subunits.

(e) a method for evaluating compounds as pharmacological modifiers of the function of the novel calcium channel subunits using the cell lines expressing those subunits alone or in combination with other calcium channel subunits.

(f) Use of the compounds identified as set forth above for the treatment of conditions which are associated with undesired calcium channel activity.

These diseases include, but are not limited to; epilepsy, migraine, ataxia, schizophrenia, hypertension, arrhythmia, angina, depression and Parkinson's disease; characterization of such associations and ultimately diagnosis of associated diseases can be carried out with probes which bind to the wild-type or defective forms of the novel calcium channels.

T-type channels in particular are responsible for rebound burst firing in central neurons and are implicated in normal brain functions such as slow-wave sleep and in neurological disorders such as epilepsy and mood disorders. They are also important in pacemaker activity in the heart, hormone secretion and fertilization, and are associated with disease states such as cardiac hypertrophy and hypertension.

As used in the specification and claims of this application, the term "T-type calcium channel" refers to a voltage-gated calcium channel having a low activation voltage, generally less than -50 mV, and most commonly less than -60 mV. T-type calcium channels also exhibit comparatively negative steady-state inactivation properties and slow deactivation kinetics. The terms " $\alpha_1$  subunit" or " $\alpha_1$  calcium channel" refer to a protein subunit of a calcium channel which is responsible for pore formation and contains the voltage sensor and binding sites for calcium channel agonists and antagonists. Such subunits may be independently functional as calcium channels or may require the presence of other subunit types for complete functionality.

As used in the specification and claims of this application, the phrase "at least partially purified" refers to DNA or protein preparations in the which the specified molecule has been separated from adjacent cellular components and molecules with which it occurs in the natural state, either by virtue of performing a physical separation process or by virtue of making the DNA or protein molecule in a non-natural environment in the first place. The term encompasses cDNA molecules and expression vectors.

In accordance with the present invention, we have identified mammalian DNA sequences which code for novel T-type calcium channel  $\alpha_1$  subunits. These subunits are believed to represent new types of  $\alpha_1$  subunits of mammalian voltage-dependent calcium channels which have been designated as types  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$ .

5 A Bacterial Artificial Chromosome (BAC) sequence (bK206c7) was identified from sequences in Sanger Genome Sequencing Center (Cambridge, U.K.) and the Washington University Genome Sequencing Center (St. Louis, MO) that contains a nucleotide sequence encoding the  $\alpha_{1I}$  subunit of human T-type calcium channel. The rationale for this identification is set forth in WO 98/38301, incorporated herein by  
10 reference. The relevant nucleotide sequence and the translated amino acid sequence containing 1854 amino acids are set forth in SEQ ID NO:17 and 18.

As described in WO 98/38031, using PCR cloning techniques to identify relevant sequences within a human brain total RNA preparation, we confirmed that the novel  $\alpha_{1I}$  calcium channel subunit is present in human brain. Subcloning of the  
15 567 nt PCR product (Seq. ID No. 19, amino acids Seq. ID No. 20) and subsequent sequencing thereof showed that this product corresponds to the derived sequence from the bK206c7 BAC genomic sequence, the nucleotide sequence of which is given as SEQ ID No. 17 (amino acid sequence Seq. ID No. 18). The same experiment was performed using a rat brain RNA preparation and resulted in recovery of a  
20 substantially identical PCR product. (SEQ ID. No. 21). The protein encoded by the rat PCR product (SEQ ID No. 22) is 96% identical to the human PCR product (Seq. ID No. 20).

These sequences, which encode a partial subunit were used as a basis for constructing full length human or rat  $\alpha_{1I}$  clones. Briefly, the subcloned  $\alpha_{1I}$  PCR  
25 product is radiolabeled by random hexamer priming according to standard methods (See, Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989) *Molecular Cloning, A Laboratory Manual*. Cold Spring Harbor Press) and used to screen commercial human brain cDNA libraries (Stratagene, La Jolla, CA). The screening of cDNA libraries follows standard methods and includes such protocols as infecting bacteria with  
30 recombinant lambda phage, immobilizing lambda DNA to nitrocellulose filters and screening under medium hybridization stringency conditions with radiolabeled probe. cDNA clones homologous to the probe are identified by autoradiography. Positive clones are purified by sequential rounds of screening.

Following this protocol, most purified cDNA's are likely to be partial sequence clones due the nature of the cDNA library synthesis. Full length clones are constructed from cDNA's which overlap in DNA sequence. Restriction enzyme sites which overlap between cDNAs are used to ligate the individual cDNA's to generate a full-length cDNA. For subsequent heterologous expression, the full-length cDNA is subcloned directly into an appropriate vertebrate expression vector, such as pcDNA-3 (Invitrogen, San Diego, CA) in which expression of the cDNA is under the control of a promoter such as the CMV major intermediate early promoter/enhancer. Other suitable expression vectors include, for example, pMT2, pRC/CMV, pcDNA3.1 and pCEP4.

Following these protocols, full length mammalian  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunit cDNAs were isolated by using the 567 base pair human fragment (Seq. ID No. 19) to screen a rat brain cDNA library. Sequencing of the recovered sequences identified the three distinct classes of calcium channel subunits which have been denominated herein as  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits. For each class of subunit, complete sequencing of the largest cDNA confirmed that it represented only a portion of the predicted calcium channel coding region. Complete sequences for the three new subunits were obtained by rescreening the rat brain cDNA library with probes derived from the partial length cDNAs to obtain overlapping segments. These segments were combined to form a complete gene by restriction digestion and ligation. The complete cDNA sequences of the rat  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits are given by Sequence ID Nos. 23, 25 and 27, respectively. Corresponding amino acid sequences are given by Sequence ID Nos. 24, 26 and 28. The same techniques are employed to recover human sequences by screening of a human or other mammalian library. Thus, for example, partial length human sequences for  $\alpha_{1G}$  and  $\alpha_{1H}$  T-type calcium channels have been recovered using the same probe (Seq. ID No. 19) and the full length rat  $\alpha_{1I}$  cDNA (Seq. ID. No. 27) has been used to recover a partial length DNA encoding a human  $\alpha_{1I}$  T-type calcium channel. The DNA and amino acid sequences for these partial length human calcium channels are given by Seq. ID Nos. 30-35. A complete coding sequence for human  $\alpha_{1G}$  was obtained and is set forth, along with the deduced amino acid sequence, in Figure 6.

Once the full length cDNA is cloned into an expression vector, the vector is then transfected into a host cell for expression. Suitable host cells include *Xenopus*



oocytes or mammalian cells such as human embryonic kidney cells as described in International Patent Publication No. WO 96/39512 which is incorporated herein by reference and Ltk cells as described in US Patent No. 5,386,025 which is incorporated herein by reference. Transfection into host cells may be accomplished by

5 microinjection, lipofection, glycerol shock, electroporation calcium phosphate or particle-mediated gene transfer. The vector may also be transfected into host cells to provide coexpression of the novel  $\alpha_1$  subunits with other subunits, such as an  $\alpha_2\delta$  subunit or a  $\gamma$  subunit.

To confirm that the three full length cDNAs (sequence ID Nos. 23, 25 and 27)

10 encoded functional calcium channels, the  $\alpha_{1G}$  and  $\alpha_{1I}$  cDNAs were transiently transfected into human embryonic kidney cells and evaluated using electrophysiological recording techniques. The results are consistent with a role of these subunits in native T-type channels in nerve, muscle and endocrine cells. Similarly, a full length clone encoding human  $\alpha_{1G}$  T-type subunit was recovered and

15 verified to have the characteristic properties of T-type channels.

The resulting cell lines expressing functional calcium channels including the novel  $\alpha_1$  subunits of the invention can be used test compounds for pharmacological activity with respect to these calcium channels. Thus, the cell lines are useful for screening compounds for pharmaceutical utility. Such screening can be carried out

20 using several available methods for evaluation of the interaction, if any, between the test compound and the calcium channel. One such method involves the binding of radiolabeled agents that interact with the calcium channel and subsequent analysis of equilibrium binding measurements including but not limited to, on rates, off rates,  $K_d$  values and competitive binding by other molecules. Another such method involves

25 the screening for the effects of compounds by electrophysiological assay whereby individual cells are impaled with a microelectrode and currents through the calcium channel are recorded before and after application of the compound of interest. Another method, high-throughput spectrophotometric assay, utilizes the loading the cell lines with a fluorescent dye sensitive to intracellular calcium concentration and

30 subsequent examination of the effects of compounds on the ability of depolarization by potassium chloride or other means to alter intracellular calcium levels. Compounds to be tested as agonists or antagonists of the novel  $\alpha_{1I}$  calcium channel subunits are combined with cells that are stably or transiently transformed with a

DNA sequence encoding the  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunits of the invention and monitored using one of these techniques.

Compounds which are shown to modulate the activity of calcium channels can then be used in pharmaceutical compositions for the treatment associated with inappropriate T-type calcium channel activity. Such conditions may also include those with inappropriate calcium channel activity in general since such activity may be modified by enhancing or decreasing T-type channel activity. Conditions appropriate for such treatment include those set forth above. The compounds identified are formulated in conventional ways as set forth in Remington's "Pharmaceutical Sciences," latest edition, Mac Publishing Co., Easton, PA. Modes of administration are those appropriate for the condition to be treated and are within the ordinary skill of the practitioner.

In addition, the regulation of expression of T-type calcium channels can be achieved by constructing expression systems encoding antisense sequences or sequences designed for triplex binding to interrupt the expression of nucleotide sequences encoding the T-type calcium channels of the invention.

DNA fragments with sequences given by SEQ ID Nos. 13-17 and 19, or polynucleotides with the complete coding sequences as given by Sequence ID Nos. 23, 25 and 27 or Figure 6, or distinctive portions thereof which do not exhibit non-discriminatory levels of homology with other types of calcium channel subunits may also be used for mapping the distribution of  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunits within a tissue sample. This method follows normal histological procedures using a nucleic acid probe, and generally involves the steps of exposing the tissue to a reagent comprising a directly or indirectly detectable label coupled to a selected DNA fragment, and detecting reagent that has bound to the tissue. Suitable labels include fluorescent labels, enzyme labels, chromophores and radio-labels.

#### Heterologous Expression of Mammalian T-type Calcium Channels in Cells

##### A. Transient Transfection in Mammalian Cells

Host cells, such as human embryonic kidney cells, HEK 293 (ATCC# CRL 1573) are grown in standard DMEM medium supplemented with 2 mM glutamine and 10% fetal bovine serum. HEK 293 cells are transfected by a standard calcium-phosphate-DNA co-precipitation method using a full-length mammalian  $\alpha_1$  T-type

calcium channel cDNA (for example, Seq. ID. No. 27) in a vertebrate expression vector (for example see Current protocols in Molecular Biology). The  $\alpha_{11}$  calcium channel cDNA may be transfected alone or in combination with other cloned subunits for mammalian calcium channels, such as  $\alpha_{2\delta}$  and  $\beta$  or  $\gamma$  subunits, and also with clones for marker proteins such the jellyfish green fluorescent protein.

Electrophysiological Recording: After an incubation period of from 24 to 72 hrs the culture medium is removed and replaced with external recording solution (see below). Whole cell patch clamp experiments are performed using an Axopatch 200B amplifier (Axon Instruments, Burlingame, CA) linked to an IBM compatible personal computer equipped with pCLAMP software. Microelectrodes are filled with 3 M CsCl and have typical resistances from 0.5 to 2.5 M ohms. The external recording solution is 2 mM BaCl<sub>2</sub>, 1 mM MgCl<sub>2</sub>, 10 mM HEPES, 40 mM TEACl, 10 mM Glucose, 92 mM CsCl, (pH 7.2). The internal pipette solution is 105 mM CsCl, 25 mM TEACl, 1 mM CaCl<sub>2</sub>, 11 mM EGTA, 10 mM HEPES (pH 7.2). Currents are typically elicited from a holding potential of -100 mV to various test potentials. Data are filtered at 1 kHz and recorded directly on the harddrive of a personal computer. Leak subtraction is carried out on-line using a standard P/5 protocol. Currents are analyzed using pCLAMP versions 5.5 and 6.0. Macroscopic current-voltage relations are fitted with the equation  $I = \frac{1}{1 + \exp(-(V_m - V_h)/S)} \times G - (V_m - E_{rev})$ , where  $V_m$  is the test potential,  $V_h$  is the voltage at which half of the channels are activated, and  $S$  reflects the steepness of the activation curve and is an indication of the effective gating charge movement. Inactivation curves are normalized to 1 and fitted with  $I = 1 / (1 + \exp((V_m - V_h)/S))$  with  $V_m$  being the holding potential. Single channel recordings are performed in the cell-attached mode with the following pipette solution (in mM): 100 BaCl<sub>2</sub>, 10 HEPES, pH 7.4 and bath solution: 100 KCl, 10 EGTA, 2 MgCl<sub>2</sub>, 10 HEPES, pH 7.4.

#### B. Transient Transfection in Xenopus Oocytes

Stage V and VI Xenopus oocytes are prepared as described by Dascal et al (1986), Expression and modulation of voltage-gated calcium channels after RNA injection into Xenopus oocytes. Science 231:1147-1150. After enzymatic dissociation with

collagenase, oocytes nuclei are microinjected with the human  $\alpha_{11}$  calcium channel cDNA expression vector construct (approximately 10 ng DNA per nucleus) using a Drummond nanoject apparatus. The  $\alpha_{11}$  calcium channel may be injected alone, or in combination with other mammalian calcium channel subunit cDNAs, such as the  $\alpha_2\text{-}\delta$  and  $\beta_1\text{b}$  and  $\gamma$  subunits. After incubation from 48 to 96 hrs macroscopic currents are recorded using a standard two microelectrode voltage-clamp (Axoclamp 2A, Axon Instruments, Burlingame, CA) in a bathing medium containing (in mM): 40 Ba(OH)<sub>2</sub>, 25 TEA-OH, 25 NaOH, 2 CsOH, 5 HEPES (pH titrated to 7.3 with methan-sulfonic acid). Pipettes of typical resistance ranging from 0.5 to 1.5 M ohms are filled with 2.8M CsCl, 0.2M CsOH, 10mM HEPES, 10mM BAPTA free acid. Endogenous Ca (and Ba) -activated Cl currents are suppressed by systematically injecting 10-30 nl of a solution containing 100mM BAPTA-free acid, 10mM HEPES (pH titrated to 7.2 with CsOH) using a third pipette connected to a pneumatic injector. Leak currents and capacitive transients are subtracted using a standard P/5 procedure.

15

#### Construction of Stable Cell Lines Expressing Mammalian T-type Calcium Channels

Mammalian cell lines stably expressing human  $\alpha_{11}$  calcium channels are constructed by transfecting the  $\alpha_{11}$  calcium channel cDNA into mammalian cells such as HEK 293 and selecting for antibiotic resistance encoded for by an expression vector. Briefly, a full-length mammalian T-type calcium channel  $\alpha_1$  subunit cDNA (for example Seq. ID No. 27) subcloned into a vertebrate expression vector with a selectable marker, such as the pcDNA3 (InvitroGen, San Diego, CA), is transfected into HEK 293 cells by calcium phosphate coprecipitation or lipofection or electroporation or other method according to well known procedures (Methods in Enzymology, Volume 185, Gene Expression Technology (1990) Edited by Goeddel, D.V.). The  $\alpha_{11}$  calcium channel may be transfected alone, or in combination with other mammalian calcium channel subunit cDNAs, such as the  $\alpha_2\text{-}\delta$  and  $\beta_1\text{b}$  subunits, either in a similar expression vector or other type of vector using different selectable markers. After incubation for 2 days in nonselective conditions, the medium is supplemented with Geneticin (G418) at a concentration of between 600 to 800  $\mu\text{g/ml}$ . After 3 to 4 weeks in this medium, cells which are resistant to G418 are visible and can be cloned as isolated colonies using standard cloning rings. After growing up

each isolated colony to confluency to establish cell lines, the expression of  $\alpha_{11}$  calcium channels can be determined at with standard gene expression methods such as Northern blotting, RNase protection and reverse-transcriptase PCR.

5 The functional detection of  $\alpha_{11}$  calcium channels in stably transfected cells can be examined electrophysiologically, such as by whole patch clamp or single channel analysis (see above). Other means of detecting functional calcium channels include the use of radiolabeled  $^{45}\text{Ca}$  uptake, fluorescence spectroscopy using calcium sensitive dyes such as FURA-2, and the binding or displacement of radiolabeled ligands that interact with the calcium channel.

10

### EXAMPLE 1

#### Partial Rat and Human Subunits

In order to recover mammalian sequences for novel calcium channels, the 567 base pair partial length human brain  $\alpha_{11}$  cDNA described in WO 98/3801 was gel-purified, radio-labelled with  $^{32}\text{P}$  dATP and dCTP by random priming (Feinberg et al., 15 1983, *Anal. Biochem.* 132: 6-13) and used to screen a rat brain cDNA library constructed in the phase vector Lambda Zapp II. (Snutch et al., 1990, *Proc Natl Acad Sci (USA)* 87: 3391-3395). Screening was carried out at 62°C in 5XSSPE (1XSSPE= 0.18 M NaCl; 1mM EDTA; 10 mM sodium phosphate, pH=7.4 0.3% SDS, 0.2 mg/ml denatured salmon sperm DNA). Filters were washed at 62°C in 0.2X SSPE/0.1% 20 SDS. After three rounds of screening and plaque purification, positive phages were transformed into Bluescript phagemids (Stratagene, La Jolla, CA) by *in vivo* excision.

Double stranded DNA sequencing on the recombinant phagemids was performed using a modified dideoxynucleotide protocol (Biggin et al., 1983, *Proc Natl Acad Sci (USA)* 80:3963-3965) and Sequenase version 2.1 (United States 25 Biochemical Corp.). DNA sequencing identified three distinct classes of calcium channel  $\alpha_1$  subunits: designated as  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{11}$  calcium channel subunits.

For each class of calcium channel  $\alpha_1$  subunit, the largest cDNA was completely sequenced and determined to represent only a portion of the predicted calcium channel coding region. In order to isolate the remaining portions of  $\alpha_{1G}$  and 30  $\alpha_{11}$  calcium channel subunits, the  $\alpha_{1G}$  clone was digested with HindIII and SpeI. The resulting 540 base pair fragment was gel purified, radiolabeled with  $^{32}\text{P}$  dATP and dCTP by random priming and used to rescreen the rat brain cDNA library as described above. The sequence of the 540 base pair  $\alpha_{1G}$  screening probe used is given

by Seq. ID No. 29. Other sequences spanning regions of distinctiveness within the sequences for the subunits could also be employed.

Double-stranded DNA sequencing of the purified recombinant phagemids showed that additional  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunit cDNAs overlapped with the original partial length cDNAs and together encoded complete protein coding regions as well as portions of their respective 5' and 3' non-coding untranslated regions.

To recover further human sequences for the novel  $\alpha_{1G}$  and  $\alpha_{1H}$  calcium channels, the 567 base pair partial length human brain  $\alpha_{1I}$  cDNA (Seq. 19) was radio-labelled with  $^{32}\text{P}$  dATP and dCTP by random priming and used to screen a commercial human thalamus cDNA library (Clontech). Hybridization was performed overnight at 65 °C in 6 X SSPE; 0.3% SDS; 5X Denhardt's. Filters were washed at 65 °C in 0.1 X SSPE/ 0.3% SDS. After four rounds of screening and plaque purification, positive phages were selected, DNA prepared and the insert cDNA excised from the lambda vector by digestion with Eco R1 restriction enzyme. The excised cDNA was subcloned into the plasmid Bluescript KS (Stratagene, La Jolla, CA) and the DNA sequence determined using a modified dideoxynucleotide protocol and Sequence version 2.1. The partial length  $\alpha_{1G}$  cDNA isolated consisted of 2212 base pairs of which 279 base pairs were 5' noncoding and 1,933 base pairs were coding region representing 644 amino acids (Seq. ID Nos. 30, 31). The partial  $\alpha_{1H}$  cDNA isolated consisted of 1,608 base pairs of which 53 base pairs were 5' noncoding and 1,555 were coding region representing 518 amino acids (Seq. ID Nos. 32, 33).

To recover further human sequences for the novel  $\alpha_{1I}$  calcium channel, the full-length rat brain  $\alpha_{1I}$  cDNA (Seq. 27; see example 2) was radio-labelled  $^{32}\text{P}$  dATP and dCTP by random priming and used to screen a commercial human hippocampus cDNA library (Stratagene). Hybridization was performed overnight at 65°C in 6 X SSPE; 0.3% SDS; 5X Denhardt's. Filters were washed at 65° C in 0.1 X SSPE/ 0.3% SDS. After four rounds of screening and plaque purification, positive phages were transformed into Bluescript phagemids (Stratagene, LA Jolla, CA) by *in vitro* excision. The excised cDNA DNA sequence was determined using a modified dideoxynucleotide protocol and Sequenase version 2.1. The partial  $\alpha_{1I}$  cDNA isolated

consisted of 1,080 base pairs of coding region representing 360 amino acids (Seq. ID Nos. 34, 35).

## EXAMPLE 2

### Full Length Rat Subunits

5 Double-stranded DNA sequencing of the purified recombinant phagemids from rat brain showed that additional  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channel cDNAs overlapped with the original partial length cDNAs and together encoded complete protein coding regions as well as portions of their respective 5' and 3' non-coding untranslated regions. (Seq. ID Nos. 23 and 27, respectively) In addition to the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium  
10 channel classes, DNA sequencing of the recombinant phagemids also identified a third class of calcium channel  $\alpha_1$  subunit: designated as the  $\alpha_{1H}$  calcium channel subunit. The partial length  $\alpha_{1H}$  calcium channel cDNAs overlapped and together encoded a complete  $\alpha_{1H}$  coding region as well as portions of the 5' and 3' untranslated regions (Seq. ID. No. 25).

15 Electrophysiological studies were performed on transiently-transfected human embryonic kidney cells (HEK-tsa201) prepared using the general protocol above. Transfection was carried out by standard calcium phosphate precipitation. (Okayama *et al.*, 1991, *Methods in Molec. Biol.*, Vol. 7, ed. Murray, E.J.). For maintenance, HEK-tsa201 cells were cultured until approximately 70% confluent, the media  
20 removed and cells dispersed with trypsin and gentle trituration. Cells were then diluted 1:10 in culture medium (10% FBS, DMEM plus L-glutamine, pen-stp) warmed to 37°C and plated onto tissue culture dishes. For transient transfection, 0.5 mM  $\text{CaCl}_2$  was mixed with a total of 20  $\mu\text{g}$  of DNA (consisting of 3 $\mu\text{g}$  of either rat brain  $\alpha_{1G}$  or  $\alpha_{1I}$  calcium channel cDNA, 2  $\mu\text{g}$  of CD8 plasmid marker, and 15  $\mu\text{g}$  of  
25 Bluescript plasmid carrier DNA). The DNA mixture was mixed thoroughly and then slowly added dropwise to 0.5 ml of 2 times HeBS (274 mM NaCl, 20mM D-glucose, 10mM KCl, 1.4 mM  $\text{Na}_2\text{HPO}_4$ , 40 mM Hepes (pH=7.05). After incubation at room temperature for 20 min, 100  $\mu\text{l}$  of the DNA mixture was slowly added to each dish of HEK-tsa201 cells and then incubated at 37°C for 24 to 48 hours in a tissue culture  
30 incubator (5%  $\text{CO}_2$ ).

Positive transfectant cells were identified visually by addition of 1  $\mu\text{l}$  of mouse CD8 (Lyt2) Dynabeads directly to the recording solution and gentle swirling to mix. Whole cell patch clamp readings were carried out with an Axopatch 200A amplifier

(Axon Instruments) as described previously. (Zamponi *et al.*, 1997, *Nature* 385: 442-446). The external recording solution was 2 mM  $\text{CaCl}_2$ , 1 mM  $\text{MgCl}_2$ , 10 mM HEPES, 40 mM TEA-Cl, 10 mM glucose, 92 mM CsCl, pH=7.2 with TEA-hydroxide. The internal pipette solutions was 105 mM CsCl, 25 mM TEA-Cl, 1mM  
5  $\text{CaCl}_2$ , 11 mM EGTA, 10 mM HEPES, pH 7.2 with NaOH.

For determination of current-voltage (I-V) relationships, cells were held at a resting potential of -100 mV and then stepped to various depolarizing test potentials. For steady-state inactivation, cells were held at various potentials for 20 sec. and currents recorded during a subsequent test pulse to the peak potential of the I-V. Leak  
10 currents and capacitative transients were subtracted using a P/5 procedure.

Figs. 1-4 show the results obtained for HEK cells transfected with and expressing the cDNA of sequences ID Nos. 23 and 27, which correspond to the subunits designated as  $\alpha_{1G}$  and  $\alpha_{1I}$ . Figs. 1A and B and 2A and B shows a comparison of the waveforms and current-voltage relationship for the two channel subunit types.  
15 In the presence of recording solution containing 2mM  $\text{Ca}^{2+}$ , both the  $\alpha_{1G}$  and  $\alpha_{1I}$  channel subunits exhibit activation properties consistent with native T-type calcium currents. Figs 1 A and 2A show the subunit calcium current from a cell held at -120 mV and depolarized to a series of test potentials. Figs 1B and 2B show the magnitude of the calcium current. From a holding potential of -110 mV, both channel first  
20 activate at approximately -70 mV and peak currents are obtained between -40 and -50 mV. Upon depolarization to various test potentials, the current waveforms of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels exhibit an overlapping pattern characteristic of native T-type channels in nerve, muscle and endocrine cells.

Fig. 3 shows steady-state inactivation profiles for the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium  
25 channels in HEK 293 cells transiently transformed with full length cDNAs (SEQ ID Nos 23 or 27) for  $\alpha_{1G}$  or  $\alpha_{1I}$  subunits. Steady state inactivation properties were determined by stepping from -120 mV to prepulse holding potentials between -120 mV and -50 mV for 15 sec., prior to a test potential of -30 mV. The data are plotted as normalized whole cell current versus prepulse holding potential and show that  $\alpha_{1G}$   
30 exhibits a  $V_{50}$  of approximately -85 mV and  $\alpha_{1I}$  a  $V_{50}$  of approximately -93 mV. Thus, consistent with native T-type calcium channels, both of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels exhibit pronounced steady-state inactivation at negative potentials.



Figs. 4A-C show a comparison of the voltage-dependent deactivation profiles of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels. HEK 293 cells were transiently transfected with either an  $\alpha_{1G}$  or  $\alpha_{1I}$  subunit cDNA (Seq. ID No. 23 or 27). The deactivation properties of  $\alpha_{1G}$  were determined by stepping from a holding potential of -100 mV to -40mV for 9 msec, and then to potentials between -120 mV and -45 mV. The deactivation properties of  $\alpha_{1I}$  were determined by stepping from a holding potential of -100 mV to -40 mV for 20 msec, and then to potentials between -120 mV and -45 mV. Both channels exhibit slow deactivation kinetics compared to typical high-threshold channels, and is consistent with the  $\alpha_{1G}$  and  $\alpha_{1I}$  subunits being subunits for T-type calcium channels

### Example 3

#### Cloning of a Full Length cDNA for the Human $\alpha_{1G}$ T-Type Calcium Channel Subunit

##### Materials and Methods:

A full length cDNA encoding the human  $\alpha_{1G}$  subunit was constructed from 5 overlapping clones (Figure 1B) isolated from a human thalamus cDNA library constructed in  $\lambda$ gt11 vector (Clontech, Cat#HL5009b).

Three  $\lambda$ gt11 cDNA clones were isolated by conventional filter hybridization.

Clone 1 was identified by hybridization to a 567 bp cDNA probe (SEQ ID NO: 19) containing the transmembrane region S4 to S6 of domain I of the previously cloned human neuronal  $\alpha_{1I}$  T-type calcium channel subunit. Clones HG10-1112 and HG5-1211 were identified by hybridization to a 1265 bp cDNA probe of the rat  $\alpha_{1H}$  T-type calcium channel subunit spanning domain II and part of the II-III intracellular loop. cDNA probes were  $^{32}$ P-dCTP labelled by random priming using a Multiprime DNA labeling system (Amersham Pharmacia). Plaque lifts using H-bond nylon membranes were done in duplicate following the standard protocols supplied by manufacturer (Amersham Pharmacia). Hybridization was performed for at least 16hrs at 65°C for clone 1 and for at least 16hrs at 58°C, clones HG10-1112 and HG5-1211. Membranes were washed in 0.1X SSC/0.3% SDS at 62°C for clone 1 and 0.2X SSC/0.1% SDS at 58°C clones HG10-1112 and HG5-1211. Blots were exposed to BioMax MS Kodak film with Kodak HE intensifying screens for at least 48hrs at -80°C. Double positive plaques were isolated and re-screened to isolate single clones

according to the procedure above. Bacteriophage DNAs were then isolated according to the  $\lambda$ gt11 library User Manual (Clontech). Clone 1 cDNA insert was excised with EcoRI (NEB) and subcloned into pBluescriptKS (Stratagene). Clones HG10-1112 and HG5-1211 cDNA inserts were excised from  $\lambda$ DNA with Not I (NEB) and subcloned into the Not I site of pBluescriptKS. Plasmids with cDNA inserts were transformed by electroporation into XL-I E.Coli host strain bacteria and sequenced using universal reverse and forward primers according to Sanger double stranded DNA sequencing method in combination with automatic sequencing ABI 100 PRISM model 377 Version3.3 (PE Biosystems).

Clone 1 was identified as a human  $\alpha_{1G}$  subunit containing the 5'UTR and 1933 bp of the in-frame coding region, including part of the intracellular I-II loop. Clone HG10-1112 was identified as a human  $\alpha_{1G}$  subunit of 3915 bp, spanning Domain I (IS5-IS6) to the III-IV loop. Clone HG5-1211 was identified as human  $\alpha_{1G}$  subunit of 3984 bp containing the I-II linker and C-terminus.

For expression in HEK cells, removal of 5' UTR from clone 1 was achieved by replacing 5'UTR DNA fragment flanked by Hind III/SacII restriction sites with 5'end - 291 bp cDNA fragment, containing translation start site and an incorporated Hind III site for subsequent cloning into pcDNA3.1 (Invitrogen). Following PCR conditions were used: 94°C -30 sec, 45°C -30 sec, 72°C -30 sec for 5 cycles and followed by 94°C -30 sec, 48°C -30 sec, 72°C -30 sec for 20 cycles (Bio-rad Gene Cyclor). The cDNA fragment was subcloned into p-Gem-T-Easy plasmid vector (Promega) and the DNA sequence determined.

The remaining region of the 3'  $\alpha_{1G}$  subunit cDNA was obtained using the PCR method on a human thalamus cDNA library with primers MD19-sense (5'GCG TGG AGC TCT TTG GAG 3') and G26- antisense (5' GCA CCC AGT GGA GAA AGG TG 3'). The PCR protocol used was 94°C -30 sec, 58°C -30 sec, 72°C -30 sec for 25 cycles (Bio-rad Gene Cyclor). A cDNA fragment of 1617 bp was subcloned into p-Gem-T-Easy plasmid vector (Promega) and sequenced. The 3'PCR cDNA was identified as a human  $\alpha_{1G}$  subunit spanning from Domain IV-S5 to the carboxyl terminus including the stop codon.

Unique restriction sites (Figures 5A and B) of the partial cDNA clones were used to construct the full length human  $\alpha_{1G}$  T-type calcium channel in pcDNA3.1 Zeo

(+) (Invitrogen) mammalian expression vector.

The complete nucleotide and amino acid sequences are shown in Figure 6.

In order to determine the functional properties of the human  $\alpha_{1G}$  channel standard calcium-phosphate transfection was used to transiently express the channel in HEK ts201 cells. Cells were cultured in Dulbecco's modified eagle medium (DMEM) supplemented with 10% fetal bovine serum, 200 U/ml penicillin and 0.2 mg/ml streptomycin at 37°C with 5% CO<sub>2</sub>. At 85% confluency cells were split with 0.25% trypsin/1 mM EDTA and plated at 10% confluency on glass coverslips. At 12 hours the medium was replaced and the cells transiently transfected using a standard calcium phosphate protocol and the  $\alpha_{1G}$  calcium channel cDNA. Fresh DMEM was supplied and the cells transferred to 28°C/5% CO<sub>2</sub>. Cells were incubated for 1 to 2 days prior to whole cell recording. Whole cell patch recordings were performed using an Axopatch 200B amplifier (Axon Instruments) linked to an IBM compatible personal computer equipped with pCLAMP version 7.0 software. The intrapipette solution contained (in mM): 105 CsCl, 25 CsCl, 1 CaCl<sub>2</sub>, 11 EGTA, 10 HEPES, pH 7.2. The extracellular solution contained (in mM): 40 TEA-Cl, 2 CaCl<sub>2</sub>, 1 MgCl<sub>2</sub>, 92 CsCl, 10 glucose, 10 HEPES, pH 7.2.

Figure 7 shows that the human  $\alpha_{1G}$  cDNA encodes a calcium channel with typical properties of a T-type current. The left panel illustrates representative current traces obtained from a holding potential of -100 mV to test pulses potentials of -90 mV to +20 mV. The traces show a typical crossover pattern and considerable inactivation during the test pulse, both of which are consistent with native T-type channels. The right panel shows a plot of the peak whole current at various test potentials and indicates that the human  $\alpha_{1G}$  cDNA first activates near -60 mV with maximal current near -40 mV, which is also consistent with native low-threshold T-type calcium channels.

Claims

1. A DNA molecule which comprises an expression cassette wherein said expression cassette comprises a nucleotide sequence encoding a T-type calcium channel  $\alpha_1$  subunit, said encoding sequence operably linked to control sequences to effect its expression.  
5
2. The DNA molecule of claim 1 wherein said  $\alpha_1$  subunit is  $\alpha_{1G}$ ,  $\alpha_{1H}$ , or  $\alpha_{1I}$ .
3. The DNA molecule of claim 2 wherein said  $\alpha_1$  subunit is derived from a mammal.
- 10 4. Recombinant host cells modified to contain the DNA molecule of any of claims 1-3.
5. The cells of claim 4 which are mammalian cells.
6. A method to effect production of a functional calcium channel which method comprises culturing the cells of claim 4 or 5 under conditions wherein said  
15 functional calcium channels are produced.
7. A method to identify a compound which is a modulator for T-type mammalian calcium channels, which method comprises contacting the cells employed in the method of claim 6 with said compound and assessing the effect of said compound on said cells.
- 20 8. A T-type calcium channel modulator identified by the method of claim 7.
9. A method to treat conditions characterized by undesirable levels of T-type calcium channel activity which method comprises administering to a subject in need of such treatment an effective amount of the modulator of claim 8.

10. The method of claim 9 wherein said condition is cardiac hypertrophy, cardiac arrhythmia, hypertension, a sleep disorder, or epilepsy.

11. A DNA molecule which comprises an expression system for a nucleotide sequence which is complementary to the nucleotide sequence encoding a T-type calcium channel  $\alpha_1$  subunit or which forms a triple helix with DNA comprising said encoding sequence.

12. A method to treat a condition characterized by an undesirable level of T-type calcium channel activity which method comprises administering to a subject in need of such treatment an effective amount of the DNA molecule of claim 11.

13. The method of claim 12 wherein said condition is cardiac hypertrophy, cardiac arrhythmia, hypertension, a sleep disorder, or epilepsy.

14. An oligonucleotide which consists essentially of a nucleotide sequence characteristic of a T-type calcium channel  $\alpha_1$  subunit, said oligonucleotide coupled to or comprising a detectable label.

15. A method to map the distribution of T-type calcium channels in a tissue which method comprises contacting said tissue with the oligonucleotide of claim 14.

16. Antibodies specifically immunoreactive with the extracellular portions of a T-type calcium channel.

17. A method to map the distribution of T-type calcium channels in a tissue which method comprises contacting said tissue with the antibodies of claim 16.

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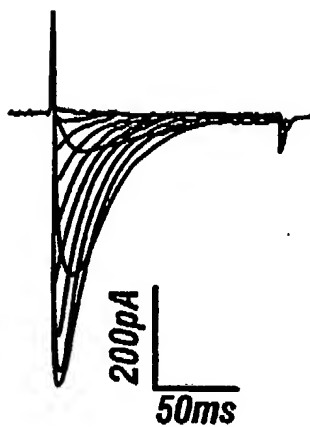


FIG. 1A

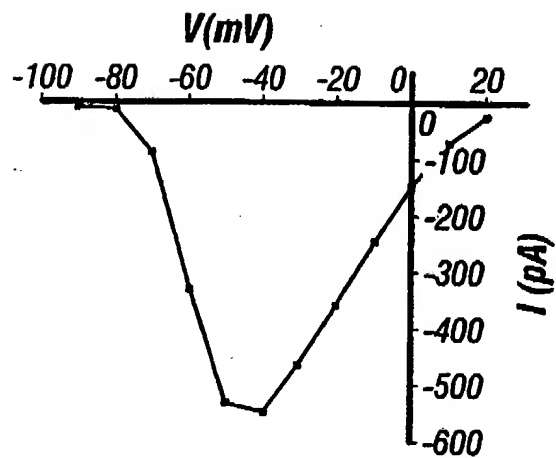


FIG. 1B

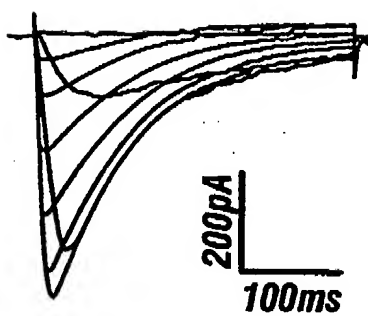


FIG. 2A

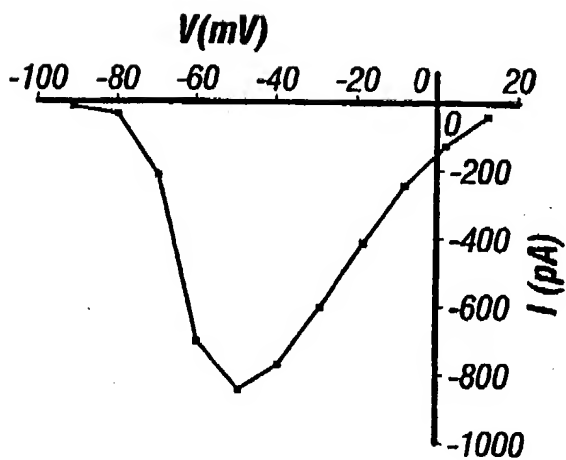


FIG. 2B

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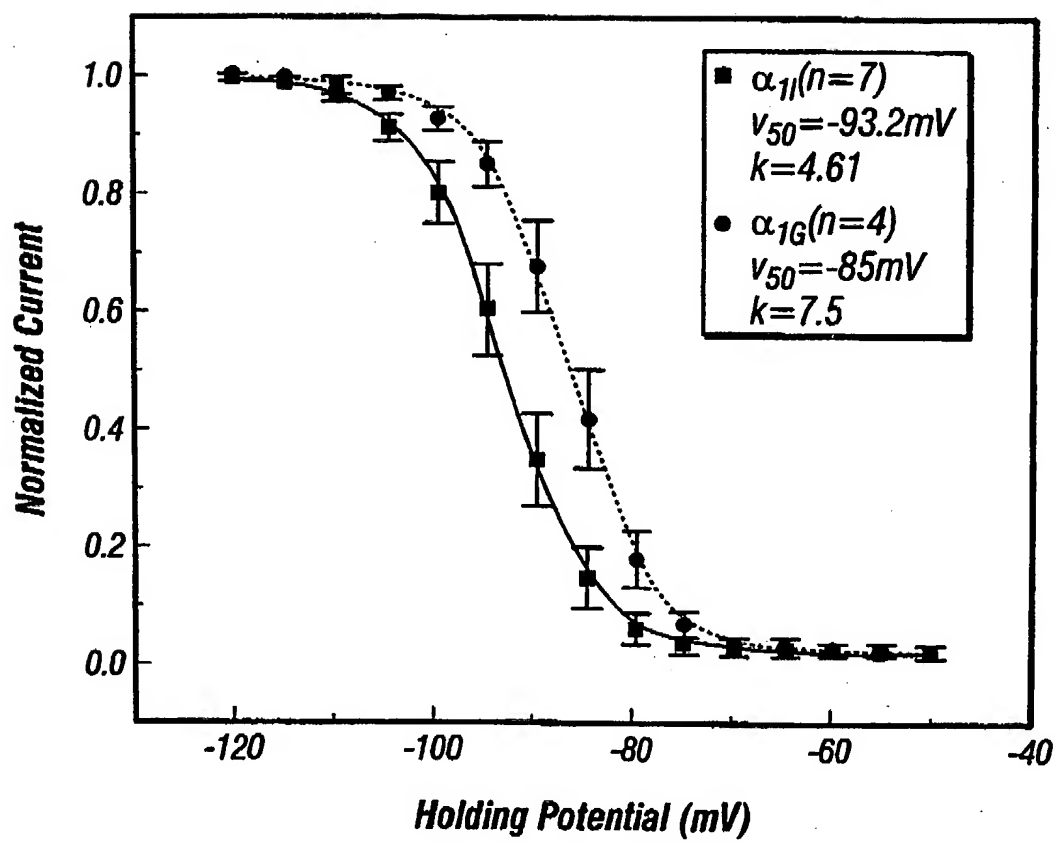


FIG. 3

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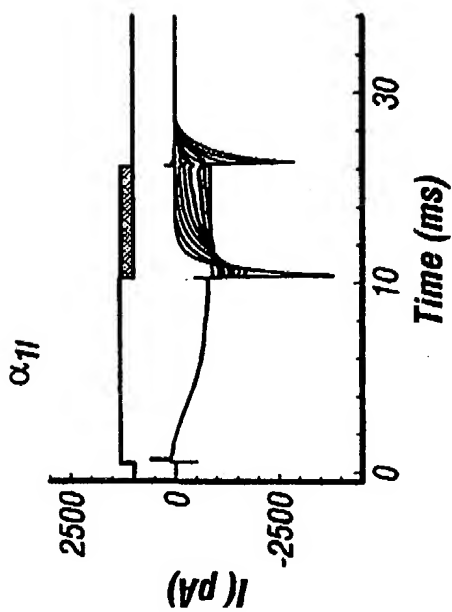


FIG. 4B

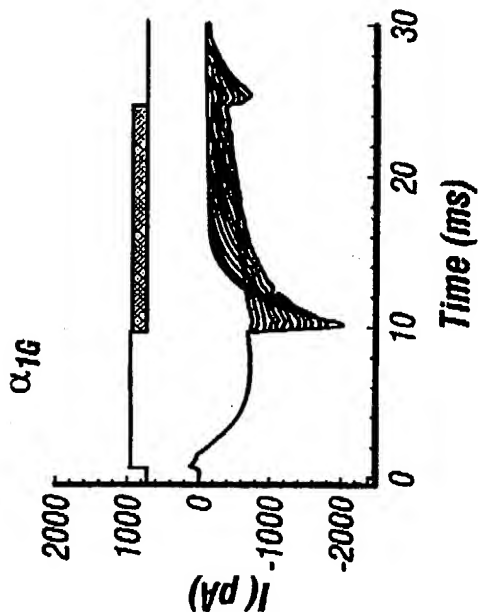


FIG. 4C

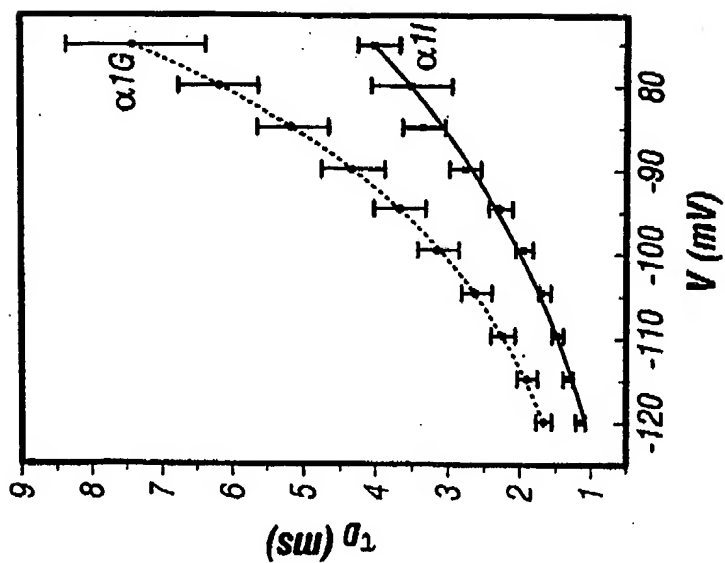


FIG. 4A



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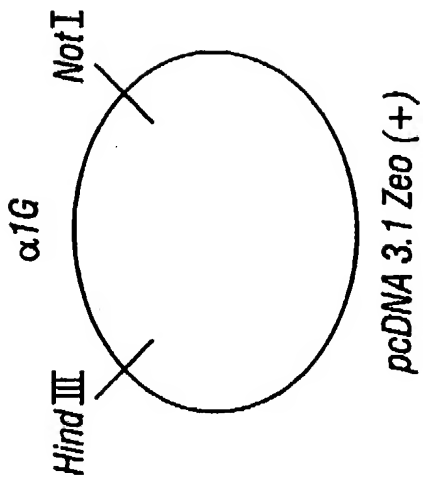
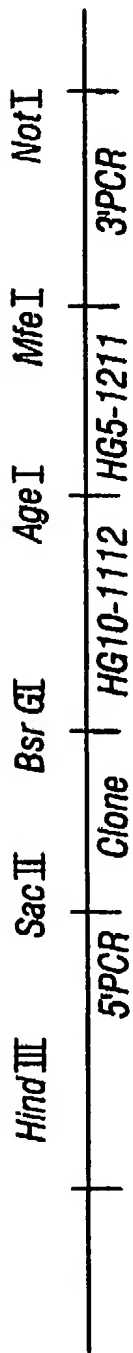


FIG. 5A

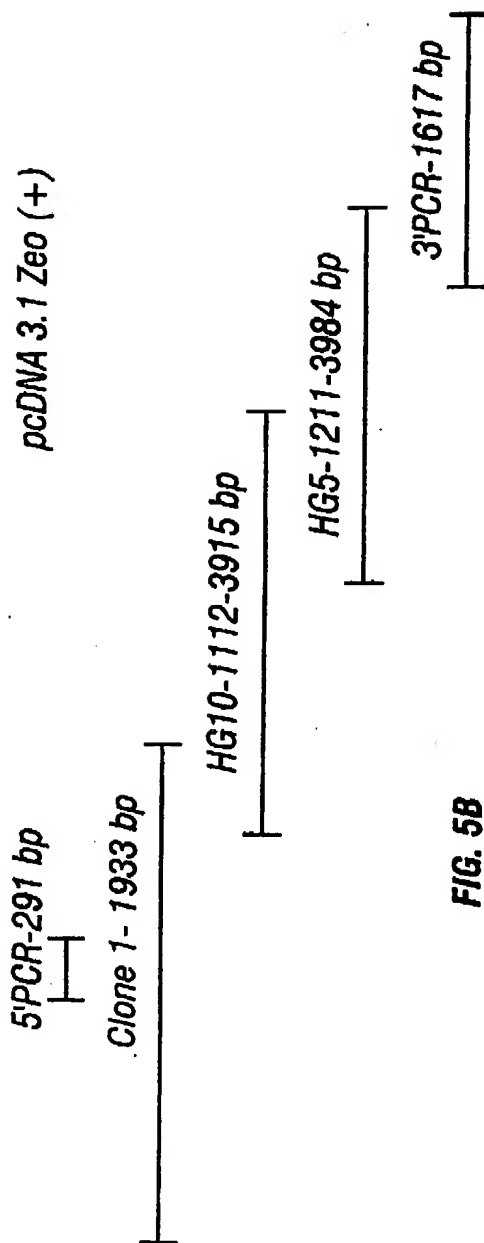


FIG. 5B

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1 aagcttgcttgccccctctccggtatcgccccggggccccggctggccagagg ATG GAC GAG GAG GAG GAT GGA 71
1      M D E E D G 7
72 GCG GGC GCC GAG GAG TCG GGA CAG CCC CGG AGC TTC ATG CCG CTC AAC GAC CTG TCG GGG 131
8 A G A E E S G Q P R S F M R L N D L S G 27
132 GCC GGG GGC CCG CCG GGG TCA GCA GAA AAG GAC CCG GGC AGC GCG GAC TCC GAG 191
28 A G G R P G P G S A E K D P G S A D S E 47
192 GCG GAG GGG CTG CCG TAC CCG GCG CTG GCC CGG GTG GTT TTC TTC TAC TTG AGC CAG GAC 251
48 A E G L P Y P A L A P V F F Y L S Q D 67
252 AGC CGC CCG AGC TGG TGT CTC CGC ACG GTC TGT AAC CCC TGG TTT GAG CGC ATC AGC 311
68 S R P R S W C L R T V C N P W F E R I S 87
312 ATG TTG GTC ATC CTT CTC AAC TGC GTG ACC CTG GGC ATG TTC CCG CCA TGC GAG GAC ATC 371
88 M L V I L L N C V T L G M F R P C E D I 107
372 GCC TGT GAC TCC CAG CGC TGC CCG ATC CTG CAG GCC TTT GAT GAC TTC ATC TTT GCC TTC 431
108 A C D S Q R C R I L Q A F D D F I F A F 127
432 TTT GCC GTG GAG ATG GTG AAG ATG GTG GCC TTG GGC ATC TTT GGG AAA AAG TGT TAC 491
128 F A V E M V V K N V A L G I F G K K C Y 147
492 CTG GGA GAC ACT TGG AAC CCG CTT GAC TTT TTC ATC GTC ATC GCA GGG ATG CTG GAG TAC 551
148 L G D T N N R L D F F I V I A G M L E Y 167
552 TCG CTG GAC CTG CAG AAC GTC AGC TTC TCA GCT GTC AGG ACA GTC CGT GTG CTG CGA CCG 611
168 S L D L Q N V S F S A V R T V R V L R P 187

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FIG. 6A

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612 CTC AGG GCC ATT AAC CGG GTG CCC AGC ATG CGC ATC CTT GTC ACG TTG CTG CTG GAT ACG 671  
 188 L R A I N R V P S M R I L V T L L L D T 207  
 672 CTG CCC ATG CTG GGC AAC GTC CTG CTG CTC TGC TTC TTT GTC TTC ATC TTC GGC ATC 731  
 208 L P M L G N V L L L C F F V F I F G I 227  
 732 GTC GGC GTC CAG CTG TGG GCA GGG CTG CTT CGG AAC CGA TGC TTC CTA CCT GAG AAT TTC 791  
 228 V G V Q L W A G L L R N R C F L P E N F 247  
 792 AGC CTC CCC CTG AGC GTG GAC CTG GAG CGC TAT TAC CAG ACA GAG AAC GAG GAT GAG AGC 851  
 248 S L P L S V D L E R Y Y O T E N E D E S 267  
 852 CCC TTC ATC TGC TCC CAG CCA CGC GAG AAC GGC ATG CGG TCC TGC AGA AGC GTG CCC ACG 911  
 268 P F I C S Q P R E N G M R S C R S V P T 287  
 912 CTG CGC GGC GAC GGG GGT GGC CCA CCT TGC GGT CTG GAC TAT GAG GCC TAC AAC AGC 971  
 288 L R G D G G G G P C G L D Y E A Y N S 307  
 972 TCC AGC AAC ACC ACC TGT GTC AAC TGG AAC CAG TAC TAC ACC AAC TGC TCA GCG GGG GAG 1031  
 308 S S K T T C V N W N Q Y Y T N C S A G E 327  
 1032 CAC AAC CCC TTC AAG GGC GCC ATC AAC TTT GAC AAC ATT GGC TAT GCC TGG ATC GCC ATC 1091  
 328 H N P F K G A I N F D N I G Y A W I A I 347  
 1092 TTC CAG GTC ATC ACG CTG GAG GGC TGG GTC GAC ATC ATG TAC TTT GTG ATG GAT GCT CAT 1151  
 348 F Q V I T L E G W V D I M Y F V M D A H 367  
 1152 TCC TTC TAC AAT TTC ATC TAC TTC ATC CTC ATC ATC GTG GGC TCC TTC TTC ATG ATC 1211  
 368 S F Y N F I Y F I L L I I V G S F F M I 387

FIG. 6B

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1212 AAC CTG TGC CTG GTG GTG ATT GCC ACG CAG TTC TCA GAG ACC AAG CAG CGG GAA AGC CAG 1271  
 388 N L C L V I A T Q F S E T K Q R E S Q 407  
 1272 CTG ATG CGG GAG CAG CGT GTG CGG TTC CTG TCC AAC GCC AGC ACC CTG GCT AGC TTC TCT 1331  
 408 L M R E Q R V R F L S N A S T L A S F S 427  
 1332 GAG CCC GGC AGC TGC TAT GAG GAG CTG CTC AAG TAC CTG GTG TAC ATC CTT CGT AAG GCA 1391  
 428 E P G S C Y E E L L K Y L V Y I L R K A 447  
 1392 GCC CGC AGG CTG GCT CAG GTC TCT CGG GCA GCA GGT GTG CGG GTT GGG CGT CTC AGC AGC 1451  
 448 A R R L A Q V S R A A G V R V G L L S S 467  
 1452 CCA GCA CCC CTC GGG GGC CAG GAG ACC CAG CCC AGC AGC AGC TGC TCT CGC TCC CAC CGC 1511  
 468 P A P L G G Q E T Q P S S S C S R S H R 487  
 1512 CGC CTA TCC GTC CAC CAC CTG GTG CAC CAC CAC CAT CAC CAC CAC TAC CAC CTG 1571  
 488 R L S V N H L V H H H N H M H N H Y H L 507  
 1572 GGC AAT GGG ACG CTC AGG GCC CCC CGG GCC AGC CCG GAG ATC CAG GAG AGG GAT GCC AAT 1631  
 508 G N G T L R A P R A S P E I Q D R D A N 527  
 1632 GGG TCC CGC AGG CTC ATG CTG CCA CCA CCC TCG ACG CCT GCC CTC TCC GGG GCC CCT 1691  
 528 G S R R L M L P P P S T P A L S G A P P 547  
 1692 GGT GGC GCA GAG TCT GTG CAC AGC TTC TAC CAT GCC GAC TGC CAC TTA GAG CCA GTC CGC 1751  
 548 G G A E S V H S F Y H A D C M L E P V R 567  
 1752 TGC CAG GCG CCC CCT CCC AGG TCC CCA TCT GAG GCA TCC GGC AGG ACT GTG GGC AGC GGG 1811  
 568 C Q A P P P R S P S E A S G R T V G S G 587

FIG. 6C

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1812 AAG GTG TAT CCC ACC GTG CAC ACC AGC CCT CCA CCG GAG ACG CTG AAG GAG AAG GCA CTA 1871  
 588 K V Y P T V N T S P P P E T L K E R A L 607  
 1872 GTA GAG GTG GCT GCC AGC TCT GGG CCC CCA ACC CTC ACC AGC CTC AAC ATC CCA CCC GGG 1931  
 608 V E V A A S S G P P T L T S L N I P P G 627  
 1932 CCC TAC AGC TCC ATG CAC AAG CTG CTG GAG ACA CAG AGT ACA GGT GCC TGC CAA AGC TCT 1991  
 628 P Y S S M H K L L E T Q S T G A C Q S S 647  
 1992 TGC AAG ATC TCC AGC CCT TGC TTG AAA GCA GAC AGT GGA GCC TGT GGT CCA GAC AGC TGC 2051  
 648 C K I S S P C L K A D S G A C G P D S C 667  
 2052 CCC TAC TGT GCC CGG GCC GCG GCA GGG GAG GTG GAG CTC GCC GAC CGT GAA ATG CCT GAC 2111  
 668 P Y C A R A G A G E V E L A D R E M P D 687  
 2112 TCA GAC AGC GAG GCA GTT TAT GAG TTC ACA CAG GAT GCC CAG CAC AGC AGC CTC CGG GAC 2171  
 688 S D S E A V Y E F T Q D A Q H S D L R D 707  
 2172 CCC CAC AGC CGG CAA CGG AGC CTG GGC CCA GAT GCA GAG CCC AGC TCT GTG CTG GCC 2231  
 708 P H S R R Q R S L G P D A E P S S V L A 727  
 2232 TTC TGG AGG CTA ATC TGT GAC ACC TTC CGA AAG ATT GTG GAC AGC AAG TAC TTT GGC CGG 2291  
 728 F W R L I C D T F R K I V D S K Y F G R 747  
 2292 GGA ATC ATG ATC GCC ATC CTG GTC AAC ACA CTC AGC ATG GGC ATC GAA TAC CAC GAG CAG 2351  
 748 G I M I A I L V N T L S M G I E Y H E Q 767  
 2352 CCC GAG GAG CTT ACC AAC GCC CTA GAA ATC AGC AAC ATC GTC TTC ACC AGC CTC TTT GCC 2411  
 768 P E E L T N A L E I S N I V F T S L F A 787

FIG. 6D

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2412 CTG GAG ATG CTG CTG AAG CTG TAT GTG TTT GGC TAC ATC AAG AAT CCC TAC 2471  
 788 L E M L L K L L V Y G P F G Y I K N P Y 807  
 2472 AAC ATC TTC GAT GGT GTC ATT GTG GTC ATC AGC GTG TGG GAG ATC GTG GGC CAG CAG GGG 2531  
 808 N I F D G V I V V I S V N E I V G Q Q G 827  
 2532 GGC GGC CTG TCG GTG CTG CGG ACC TTC CGC CTG ATG CGT GTG CTG AAG CTG GTG CGC TTC 2591  
 828 G G L S V L R T F R L M R V L K L V R F 847  
 2592 CTG CCG GCG CTG CAG CGG CAG CTG GTG CTG ATG AAG ACC ATG GAC AAC GTG GCC ACC 2651  
 848 L P A L Q R Q L V V L M K T M D N V A T 867  
 2652 TTC TGC ATG CTG CTT ATG CTC TTC ATC TTC AGC ATC CTG GGC ATG CAT CTC TTC 2711  
 868 F C M L L M L F I F S I L G M H L F 887  
 2712 GGC TGC AAG TTT GCC TCT GAG CGG GAT GGG GAC ACC CTG CCA GAC CGG AAG AAT TTT GAC 2771  
 888 G C K F A S E R D G D T L P D R K N F D 907  
 2772 TCC TTG CTC TGG GCC ATC GTC ACT GTC TTT CAG ATC CTG ACC CAG GAC GAC TGG AAC AAA 2831  
 908 S L L W A I V T V F Q I L T Q E D W N K 927  
 2832 GTC CTC TAC AAT GGT ATG GCC TCC ACG TCG TCC TGG GCG GCC CTT TAT TTC ATT GCC CTC 2891  
 928 V L Y N G M A S T S S W A A L Y F I A L 947  
 2892 ATG ACC TTC GGC AAC TAC GTG CTC TTC AAT TTG CTG GTC GCC ATT CTG GTG GAG GGC TTC 2951  
 948 M T F G N Y V L F N L L V A I L V E G F 967  
 2952 CAG GCG GAG GAA ATC AGC AAA CGG GAA GAT GCG AGT GGA CAG TTA AGC TGT ATT CAG CTG 3011  
 968 Q A E E I S K R E D A S G Q L S C I Q L 987

FIG. 6E

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3012 CCT GTC GAC TCC CAG GGG GGA GAT GCC AAC AAG TCC GAA TCA GAG CCC GAT TTC TTC TCA 3071  
 988 P V D S Q G G D A N K S E S E P D F F S 1007  
 3072 CCC AGC CTG GAT GGT GAT GGG GAC AGG AAG AAG TGC TTG GCC TTG GTG TCC CTG GGA GAG 3131  
 1008 P S L D G D G D R K K C L A L V S L G E 1027  
 3132 CAC CCG GAG CTG CCG AAG AGC CTG CTG CCG CCT CTC ATC ATC CAC ACG GCC GCC ACA CCC 3191  
 1028 H P E L R K S L L P P L I I H T A A T P 1047  
 3192 ATG TCG CTG CCC AAG AGC ACC AGC GGC CTG GGC GAG GCG CTG GGC CCT GCG TCG CGC 3251  
 1048 M S L P K S T S T G L G E A L G P A S R 1067  
 3252 CGC ACC AGC AGC GGG TCG GCA GAG CCT GGG GCG GCC CAC GAG ATG AAG TCA CCG CCC 3311  
 1068 R T S S S G S A E P G A A H E M K S P P 1087  
 3312 AGC GCC CGC AGC TCT CCG CAC AGC CCC TGG AGC GCT GCA AGC AGC TGG ACC AGC AGG CGC 3371  
 1088 S A R S S P H S P W S A A S S W T S R R 1107  
 3372 TCC AGC CGG AAC AGC CTC GGC CGT GCA CCC AGC CTG AAG CCG AGA AGC CCA AGT GGA GAG 3431  
 1108 S S R N S L G R A P S L K R R S P S G E 1127  
 3432 CGG CGG TCC CTG TTG TCG GGA GAA GGC CAG GAG AGC CAG GAT GAA GAG AGC TCA GAA 3491  
 1128 R S L L S G E G Q E S Q D E E S S E 1147  
 3492 GAG GAG CGG GCC AGC CCT GCG GGC AGT GAC CAT CGC CAC AGG GGG TCC CTG GAG CGG GAG 3551  
 1148 E E R A S P A G S D H R N R G S L E R E 1167  
 3552 GCC AAG AGT TCC TTT GAC CTG CCA GAC ACA CTG CAG CTG CCA GGG CTG CAT CGC ACT GCC 3611  
 1168 A K S S F D L P D T L Q V P G L H R T A 1187

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3612 AGT GGC CGA GGG TCT GCT TCT GAG CAC CAG GAC TGC AAT CGC AAG TCG GCT TCA GGG CGC 3671  
 1188 S G R G S A S E H Q D C N G K S A S G R 1207  
 3672 CTG GCC CGG GCC CTG CGG CCT GAT GAC CCC CCA CTG GAT GGG GAT GAC GCC GAT GAC GAG 3731  
 1208 L A R A L R P D D P P P L D G D D A D D E 1227  
 3732 GGC AAC CTG AGC AAA GGG GAA CGG GTC CGC GCG TGG ATC CGA GCC CGA CTC CCT GCC TGC 3791  
 1228 G N L S K G E R V R A W I R A R L P A C 1247  
 3792 TAC CTC GAG CGA GAC TCC TGG TCA CCC TAC ATC TTC CCT CCT CAG TCC AGG TTC CGC CTC 3851  
 1248 Y L E R D S W S A Y I F P P Q S R F R L 1267  
 3852 CTG TGT CAC CGG ATC ATC ACC CAC AAG ATG TTC GAC CAG GTG GTC CTT GTC ATC ATC TTC 3911  
 1268 L C H R I I T N K M F D H V V L V I I F 1287  
 3912 CTT AAC TGC ATC ACC ATC GCC ATG GAG CGC CCC AAA ATT GAC CCC CAC AGC GCT GAA CGC 3971  
 1288 L N C I T I A M E R P K I D P H S A E R 1307  
 3972 ATC TTC CTG ACC CTC TCC AAT TAC ATC TTC ACC GCA GTC TTT CTG GCT GAA ATG ACA GTG 4031  
 1308 I F L T L S N Y I F T A V F L A E M T V 1327  
 4032 AAG GTG GCA CTG GGC TGG TGC TGC GAG CAG GCG TAC CTG CGG AGC AGT TGG AAC 4091  
 1328 K V V A L G W C F G E Q A Y L R S S W N 1347  
 4092 GTG CTG GAC GGG CTG TTG GTG CTC ATC TCC GTC ATC GAC ATT CTG GTG TCC ATG GTC TCT 4151  
 1348 V L D G L L V L I S V I D I L V S M V S 1367  
 4152 GAC AGC ACC AAG ATC CTG GGC ATG CTG AGG GTG CTG CGG CTG CGG ACC CTG CGC 4211  
 1368 D S G T K I L G M L R V L R L L R T L R 1387

FIG. 6G



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4212	CCG	CTC	AGG	GTG	ATC	AGC	CGG	GCG	CAG	GGG	CTG	AAG	CTG	GTG	GTG	GAG	ACG	CTG	ATG	TCC	4271
1388	P	L	R	V	I	S	R	A	Q	G	L	K	L	V	V	E	T	L	M	S	1407
4272	TCA	CTG	AAA	CCC	ATC	GGC	AAC	ATT	GTA	GTC	ATC	TGC	TGT	GCC	TTC	TTC	ATC	ATT	TTC	GGC	4331
1408	S	L	K	P	I	G	N	I	V	V	I	C	C	A	F	F	I	I	F	G	1427
4332	ATC	TTG	GGG	GTG	CAG	CTC	TTC	AAA	GGG	AAG	TTT	TTC	GTG	TGC	CAG	GGC	GAG	GAT	ACC	AGG	4391
1428	I	L	G	V	Q	L	F	K	G	K	F	F	V	C	Q	G	E	D	T	R	1447
4392	AAC	ATC	ACC	AAT	AAA	TCG	GAC	TGT	GCC	GAG	GCC	AGT	TAC	CGG	TGG	GTC	CGG	CAC	AAG	TAC	4451
1448	N	I	T	N	K	S	D	C	A	E	A	S	Y	R	W	V	R	M	K	Y	1467
4452	AAC	TTT	GAC	AAC	CTT	GGC	CAG	GCC	CTG	ATG	TCC	CTG	TTC	GTT	TTG	GCC	TCC	AAG	GAT	GGT	4511
1468	N	F	D	N	L	G	Q	A	L	M	S	L	F	V	L	A	S	K	D	G	1487
4512	TGG	GTG	GAC	ATC	ATG	TAC	GAT	GGG	CTG	GAT	GCT	GTG	GGC	GTG	GAC	CAG	CAG	CCC	ATC	ATG	4571
1488	W	V	D	I	M	Y	D	G	L	D	A	V	G	V	D	Q	Q	P	I	M	1507
4572	AAC	CAC	AAC	CCC	TGG	ATG	CTG	CTG	TAC	TTC	ATC	TCG	TTC	CTG	CTC	ATT	GTG	GCC	TTC	TTT	4631
1508	N	H	N	P	N	M	L	L	Y	F	I	S	F	L	L	I	V	A	F	F	1527
4632	GTC	CTG	AAC	ATG	TTT	GTG	GGT	GTG	GTG	GTG	GAG	AAC	TTC	CAC	AAG	TGT	AGG	CAG	CAC	CAG	4691
1528	V	L	N	M	F	V	G	V	V	V	E	N	F	H	K	C	R	Q	H	Q	1547
4692	GAG	GAA	GAG	GAG	GCC	CGG	CGG	CGG	GAG	GAG	AAG	CGC	CTA	CGA	AGA	CTG	GAG	AAA	AAG	AGA	4751
1548	E	E	E	E	A	R	R	R	E	E	K	R	L	R	R	L	E	K	K	R	1567
4752	AGG	AAA	GCC	CAG	TGC	AAA	CCT	TAC	TAC	TCC	GAC	TAC	TCC	CGC	TTC	CGG	CTC	CTC	GTC	CAC	4811
1568	R	K	A	Q	C	K	P	Y	Y	S	D	Y	S	R	F	R	L	L	V	H	1587

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FIG. 6H

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4812 CAC TTG TGC ACC AGC CAC TAC CTG GAC CTC TTC ATC ACA GGT GTC ATC GGG CTG AAC GTG 4871  
 1588 H L C T S H Y L D L F I T G V I G L N V 1607  
 4872 GTC ACC ATG GCC ATG GAG CAC TAC CAG CAG CCC CAG ATT CTG GAT GAG GCT CTG AAG ATC 4931  
 1608 V T M A M E H Y Q Q P Q I L D E A L K I 1627  
 4932 TGC AAC TAC ATC TTC ACT GTC ATC TTT GTC TTG GAG TCA GTT TTC AAA CTT GTG GCC TTT 4991  
 1628 C N Y I F T V I F V L E S V F K L V A F 1647  
 4992 GGT TTC CGT CGG TTC TTC CAG GAC AGG TGG AAC CAG CTG GAC CTG GCC ATT GTG CTG CTG 5051  
 1648 G F R R F F Q D R W N Q L D L A I V L L 1667  
 5052 TCC ATC ATG GGC ATC ACG CTG GAG GAA ATC GAG GTC AAC GCC TCG CTG CCC ATC AAC CCC 5111  
 1668 S I M G I T L E E I E V N A S L P I N P 1687  
 5112 ACC ATC ATC CGC ATC ATG AGG GTG CTG CGC ATT GCC CGA GTG CTG AAG CTG CTG AAG ATG 5171  
 1688 T I I R I M R V L R I A R V L K L L K M 1707  
 5172 GCT GTG GGC ATG CGG GCG CTG GAC ACG GTG ATG CAG GCC CTG CCC CAG GTG GGG AAC 5231  
 1708 A V G M R A L L D T V M Q A L P Q V G N 1727  
 5232 CTG GGA CTT CTC TTC ATG TTG TTG TTT TTC ATC TTT GCA GCT CTG GGC GTG GAG CTC TTT 5291  
 1728 L G L L F M L L F F I F A A L G V E L F 1747  
 5292 GGA GAC CTG GAG TGT GAC GAG ACA CAC CCC TGT GAG GGC CTG GGC CGT CAT GCC ACC TTT 5351  
 1748 G D L E C D E T H P C E G L G R H A T F 1767  
 5352 CGG AAC TTT GGC ATG GCC TTC CTA ACC CTC TTC CGA GTC TCC ACA GGT GAC AAT TGG AAT 5411  
 1768 R N F G M A F L T L F R V S T G D N W E 1787

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5412 GGC ATT ATG AAG GAC ACC CTC CGG GAC TGT GAC CAG GAG TCC ACC TGC TAC AAC ACG GTC 5471  
 1788 G I M K D T L R D C D Q E S T C Y F T V 1807  
 5472 ATC TCG CCT ATC TAC TTT GTG TTC GTG CTG ACG GCC CAG TTC GTG CTA GTC AAC GTG 5531  
 1808 I S P I Y F V S F V L T A Q F V L V M V 1827  
 5532 GTG ATC GCC GTG CTG ATG AAG CAC CTG GAG GAG AGC AAC AAG GAG GCC AAG GAG GAG GCC 5591  
 1828 V I A V L M K H L E E S N K E A K E E A 1847  
 5592 GAG CTA GAG GCT GAG CTG GAG ATG AAG ACC CTC AGC CCC CAG CCC CAC TCG CCA 5651  
 1848 E L E A E L E M K T L S P Q P H S P 1867  
 5652 CTG GGC AGC CCC TTC CTC TGG CCT GGG GTC GAG GGC CCC GAC AGC CCC AAG 5711  
 1868 L G S P F L N P G V E G P D S P D S P K 1887  
 5712 CCT GGG GCT CTG CAC CCA GCG GCC CAC GCG AGA TCA GCC TCC CAC TTT TCC CTG GAG CAC 5771  
 1888 P G A L M P A A H A R S A S H F S L E H 1907  
 5772 CCC ACG ATG CAG CCC CAC CCC ACG GAG CTG CCA GGA CCA GAC TTA CTG ACT GTG CGG AAG 5831  
 1908 P T M Q P P H P T E L P G P D L L T V R K 1927  
 5832 TCT GGG GTC AGC CGA ACG CAC TCT CTG CCC AAT GAC AGC TAC ATG TGT CGG CAT GGG AGC 5891  
 1928 S G V S R T M S L P N D S Y M C R G S 1947  
 5892 ACT GCC GAG GGG CCC CTG GGA CAC AGG GGC TGG GGC CTC CCC AAA GCT CAG TCA GGC TCC 5951  
 1948 T A E G P L G H R G W G L P K A Q S G S 1967  
 5952 GTC TTG TCC GTT CAC TCC CAG CCA GCA GAT ACC AGC TAC ATC CTG CAG CTT CCC AAA GAT 6011  
 1968 V L S V H S Q P A D T S Y I L Q L P K D 1987

FIG. 6J

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6012	GCA	CCT	CAT	CTG	CTC	CAG	CCC	CAC	AGC	GCC	CCA	ACC	TGG	GGC	ACC	ATC	CCC	AAA	CTG	CCC	6071
1988	A	P	H	L	L	Q	P	M	S	A	P	T	W	G	T	I	P	K	L	P	2007
6072	CCA	CCA	GGA	CGC	TCC	CCT	TTG	GCT	CAG	AGG	CCA	CTC	AGG	CGC	CAG	GCA	GCA	ATA	AGG	ACT	6131
2008	P	P	G	R	S	P	L	A	Q	R	P	L	R	R	Q	A	A	I	R	T	2027
6132	GAC	TCC	TTG	GAC	GTT	CAG	GGT	CTG	GGC	AGC	CGG	GAA	GAC	CTG	CTG	GCA	GAG	GTG	AGT	GGG	6191
2028	D	S	L	D	V	Q	G	L	G	S	R	E	D	L	L	A	E	V	S	G	2047
6192	CCC	TCC	CCG	CCC	CTG	GCC	CGG	GCC	TAC	TCT	TTC	TGG	GGC	CAG	TCA	AGT	ACC	CAG	GCA	CAG	6251
2048	P	S	P	P	L	A	R	A	Y	S	F	W	G	Q	S	S	T	Q	A	Q	2067
6252	CAG	CAC	TCC	CGC	AGC	CAC	AGC	AAG	ATC	TCC	AAG	CAC	ATG	ACC	CCG	CCA	GCC	CCT	TGC	CCA	6311
2068	Q	H	S	R	S	H	S	K	I	S	K	H	H	T	P	P	A	P	C	A	2087
6312	GGC	CCA	GAA	CCC	AAC	TGG	GGC	AAG	GGC	CCT	CCA	GAG	ACC	AGA	AGC	AGC	TTA	GAG	TTG	GAC	6371
2088	G	P	E	P	N	M	G	K	G	P	P	E	T	R	S	S	L	E	L	D	2107
6372	ACG	GAG	CTG	AGC	TGG	ATT	TCA	GGA	GAC	CTC	CTG	CCC	CCT	GGC	GGC	CAG	GAG	GAG	CCC	CCA	6431
2108	T	E	L	S	N	I	S	G	D	L	L	P	P	G	G	Q	E	E	P	P	2127
6432	TCC	CCA	CGG	GAC	CTG	AAG	AAG	TGC	TAC	AGC	GTG	GAG	GCC	CAG	AGC	TGC	CAG	CGC	CGG	CCT	6491
2128	S	P	R	D	L	K	C	Y	S	S	V	E	A	Q	S	C	Q	R	R	P	2147
6492	ACG	TCC	TGG	CTG	GAT	GAG	CAG	AGG	AGA	CAC	TCT	ATC	GCC	GTC	AGC	TGC	CTG	GAC	AGC	GGC	6551
2148	T	S	W	L	D	E	Q	R	R	H	S	I	A	V	S	C	L	D	S	G	2167
6552	TCC	CAA	CCC	CAC	CTG	GGC	ACA	GAC	CCC	TCT	AAC	CTT	GGG	GGC	CAG	CCT	CTT	GGG	GGG	CCT	6611
2168	S	Q	P	H	L	G	T	D	P	S	N	L	G	G	Q	P	L	G	G	P	2187

FIG. 6K

16/18

6612 GGG AGC CGG CCC AAG AAA AAA CTC AGC CCG CCT AGT ATC ACC ATA GAC CCC CCC GAG AGC 6671  
 2188 G S R P K K L S P P S I T I D P P E S 2207  
 6672 CAA GGT CCT CGG ACC CCG CCC AGC CCT GGT ATC TGC CTC CGG AGG AGG GCT CCG TCC AGC 6731  
 2208 Q G P R T P P S P G I C L R R A P S S 2227  
 6732 GAC TCC AAG GAT CCC TTG GCC TCT GGC CCC CCT GAC AGC ATG GCT GCC TCG CCC TCC CCA 6791  
 2228 D S K D P L A S G P P D S M A A S P S P 2247  
 6792 AAG AAA GAT GTG CTG AGT CTC TCC GGT TTA TCC TCT GAC CCA GCA GAC CTG GAC CCC TGA 6851  
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FIG. 6L

17/18

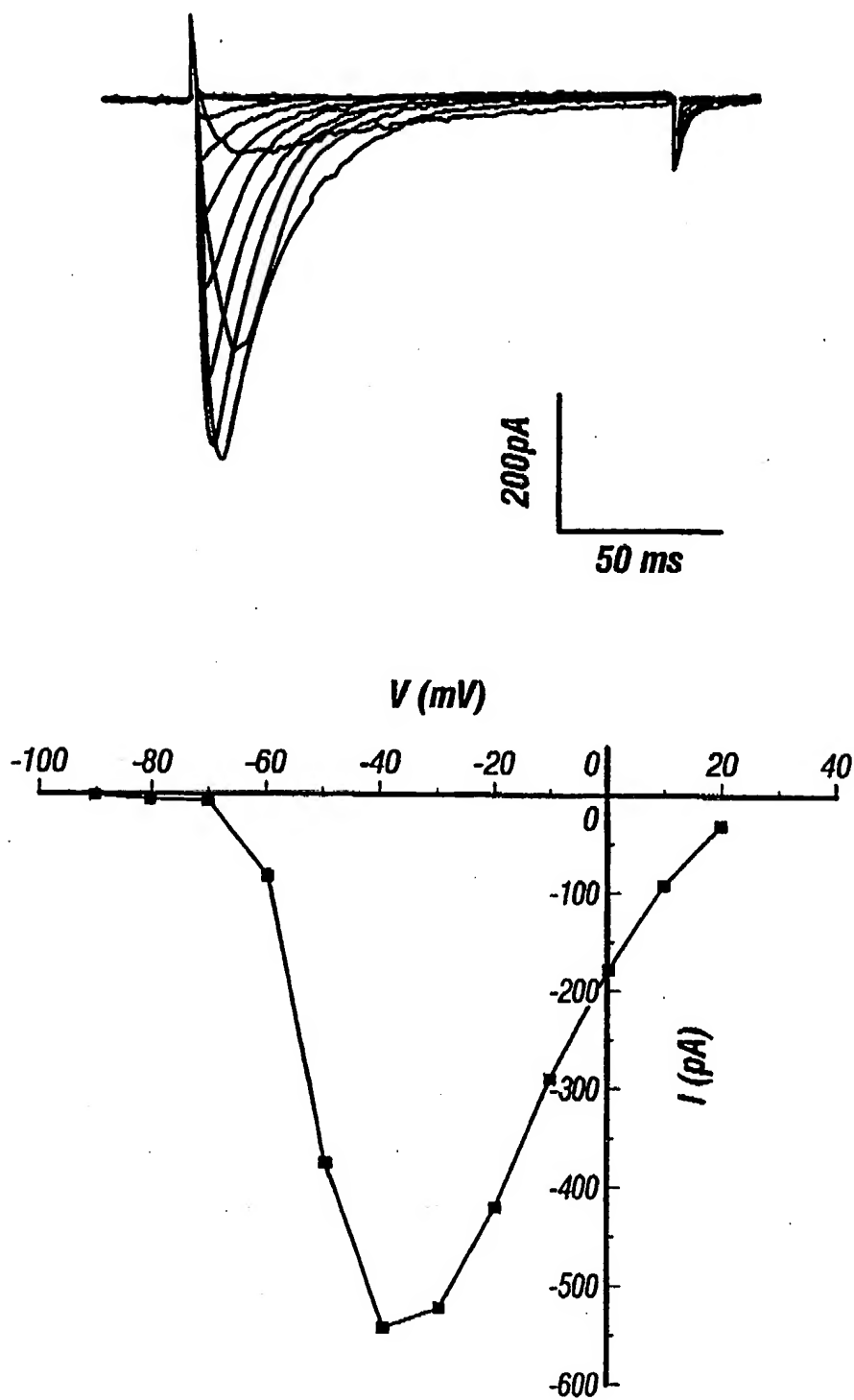


FIG. 7

SUBSTITUTE SHEET (RULE 26)

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I	II	III	IV	
LAASE E GWVYV	QIITQ E GWTDF	ETLSF K GWNVI	RCLTG E DWNDI	NIC-1 (C11D2.6)
LAASQ E GWVYV	QIITQ E GWTDV	ETLSY K GWNVV	RSVTG E DWNDI	NIC-2 (C27F2.3)
EASSQ E GWVFL	QILTQ E GWVDV	EVLSL K GWVEV	RIVTG E DWNKI	Rat -NIC
QCITM E GWTDV	QILTQ E DWNSV	TVSTF K GWPEL	RCATG E AWQDI	L-Type Ca Channel
QVITL E GWVDI	QILTQ E DWNKV	VLASK D GWVDI	RVSTG D NWNKI	T-Type Ca Channel
RLMTQ D FWENL	RVLCG E WIETM	QVATF K GWMDI	QITTS A GWDGL	Na Channels

FIG. 8

## SEQUENCE LISTING

<110> Snutch, Terry P.  
Baillie, David L.

<120> NOVEL HUMAN CALCIUM CHANNELS AND RELATED PROBES, CELL  
LINES AND METHODS

<130> NMED.P-001-2 (CIP)

<140>

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<150> 09/030,482

<151> 1998-02-25

<150> 60/039,204

<151> 1997-02-28

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<170> PatentIn Ver. 2.0

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Leu Asn Val Ala Asn Met Asp Asn Phe Phe Ala Pro Val Phe Thr Met
      20                      25                      30

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35

40

45

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 Glu Cys Val Ser Met Leu Val Ile Leu Leu Asn Cys Val Thr Leu Gly  
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Glu Thr Gly Glu Glu Pro His Ser Trp Ser Pro Arg Ala Thr Arg Arg 515 520 525		
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Pro Gly Pro His Ala Lys Glu Pro Arg His Tyr Pro Leu Thr Val Trp			
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	690	695	700
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805

810

815

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850 855 860

Ser Pro Gly Gln Gly Val Leu Ser His Pro Val Thr Pro Pro His Thr  
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Ala Pro Trp Arg Met Glu Thr Gly Lys Gln Gly His Gly Cys Glu Glu  
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Gln Ala Glu Val Thr Val Val Leu Ala Glu Glu Ala Pro Pro Gln Gly 1090 1095 1100		
Leu Arg Lys Thr Gly Arg Gly Arg Gly Gly Leu Asp Gly Gly Gly Leu 1105 1110 1115 1120		
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Asp Glu Val Gly Asp Ala Asn Arg Ser Tyr Ser Asp Glu Asp Gln Ser 1140 1145 1150		
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Tyr Asp Gln Arg Ser Leu Val Gly Gly Leu Arg Ala Thr Ala Gly Val 1235 1240 1245		
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Gly Ala Asp Pro Asn Gly Asn Ser Phe Gln Ser Ser Ser Arg Ser Ser 1265 1270 1275 1280		
Tyr Tyr Gly Pro Trp Gly Arg Ser Ala Ala Trp Ala Ser Arg Arg Ser 1285 1290 1295		
Ser Trp Asn Ser Leu Lys His Lys Pro Pro Ser Ala Glu His Glu Ser 1300 1305 1310		
Leu Leu Ser Ala Glu Arg Gly Gly Gly Ala Arg Val Cys Glu Val Ala		

1315	1320	1325
Ala Asp Glu Gly Pro Pro Arg Ala Ala Pro Leu His Thr Pro His Ala 1330	1335	1340
His His Val His His Gly Pro His Leu Ala His Arg His Arg His His 1345	1350	1355 1360
Arg Arg Thr Leu Ser Leu Asp Asn Arg Asp Ser Val Asp Leu Ala Glu 1365	1370	1375
Leu Val Pro Ala Val Gly Ala His Pro Arg Ala Ala Trp Arg Ala Ala 1380	1385	1390
Gly Pro Ala Pro Gly His Glu Asp Cys Asn Gly Arg Met Pro Ser Ile 1395	1400	1405
Ala Lys Asp Val Phe Thr Lys Met Gly Asp Arg Gly Asp Arg Gly Glu 1410	1415	1420
Asp Glu Glu Glu Ile Asp Tyr Val Ser Gly Gly Gly Ala Glu Gly Asp 1425	1430	1435 1440
Leu Thr Leu Cys Phe Arg Val Arg Lys Met Ile Asp Val Tyr Lys Pro 1445	1450	1455
Asp Trp Cys Glu Val Arg Glu Asp Trp Ser Val Tyr Leu Phe Ser Pro 1460	1465	1470
Glu Asn Arg Leu Arg Asp Leu Gly Trp Val Ser Leu Glu Cys Gln Gly 1475	1480	1485
Lys Val Gly Asp Leu Val Val Trp Val Tyr Gly Gln Arg Arg Gln Arg 1490	1495	1500
Gln Thr Ile Ile Ala His Lys Leu Phe Asp Tyr Val Val Leu Ala Phe 1505	1510	1515 1520
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Ala Gly Ser Thr Glu Arg Ile Phe Leu Thr Val Ser Asn Tyr Ile Phe 1540	1545	1550
Thr Ala Ile Phe Val Gly Glu Met Thr Leu Lys Val Val Ser Leu Gly 1555	1560	1565
Leu Tyr Phe Gly Glu Gln Ala Tyr Leu Arg Ser Ser Trp Asn Val Leu		

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Asp Gly Phe Leu Val Phe Val Ser Ile Ile Asp Ile Val Val Ser Leu 1585	1590	1595 1600
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Leu Lys Leu Val Val Glu Thr Leu Ile Ser Ser Leu Lys Pro Ile Gly 1635	1640	1645
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Gly Val Gln Leu Phe Lys Gly Lys Phe Tyr His Cys Leu Gly Val Asp 1665	1670	1675 1680
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Trp Val His His Lys Tyr Asn Phe Asp Asn Leu Gly Gln Ala Leu Met 1700	1705	1710
Ser Leu Phe Val Leu Ala Ser Lys Asp Gly Trp Val Asn Ile Met Tyr 1715	1720	1725
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Asn Pro Trp Met Leu Leu Tyr Phe Ile Ser Phe Leu Leu Ile Val Ser 1745	1750	1755 1760
Phe Phe Val Leu Asn Met Phe Val Gly Val Val Val Glu Asn Phe His 1765	1770	1775
Lys Cys Arg Gln His Gln Glu Ala Glu Glu Ala Arg Arg Arg Glu Glu 1780	1785	1790
Lys Arg Leu Arg Arg Leu Glu Lys Lys Arg Arg Lys Ala Gln Arg Leu 1795	1800	1805
Pro Tyr Tyr Ala Thr Tyr Cys His Thr Arg Leu Leu Ile His Ser Met 1810	1815	1820
Cys Thr Ser His Tyr Leu Asp Ile Phe Ile Thr Phe Ile Ile Cys Leu		

1825

1830

1835

1840

Asn Val Val Thr Met Ser Leu Glu His Tyr Asn Gln Pro Thr  
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 Gly Val Gln Leu Trp Ala Gly Leu Leu Arg Asn Arg Cys Phe Leu Glu  
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 Glu Asn Phe Thr Ile Gln Gly Asp Val Ala Leu Pro Pro Tyr Tyr Gln  
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 Pro Glu Glu Asp Asp Glu Met Pro Phe Ile Cys Ser Leu Ser Gly Asp

65

70

75

80

Asn Gly Ile Met Gly Cys His Glu Ile Pro Pro Leu Lys Glu Gln Gly  
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Arg Glu Cys Cys Leu Ser Lys Asp Asp Val Tyr Asp Phe Gly Ala Gly  
100 105 110

Arg Gln Asp Leu Asn Ala Ser Gly Leu Cys Val Asn Trp Asn Arg Tyr  
115 120 125

Tyr Asn Val Cys Arg Thr Gly Ser Ala Asn Pro His Lys Gly Ala Ile  
130 135 140

Ser Phe Asp Asn Ile Gly Tyr Ala Trp Ile Val Ile Phe Gln Val Ile  
145 150 155 160

Thr Leu Glu Gly Trp Val Ala Ile Met Tyr Tyr Val Met Asp Ala Leu  
165 170 175

Ser Phe Tyr Asn Phe Val Tyr Phe Ile Leu Leu Ile Ile  
180 185

&lt;210&gt; 21

&lt;211&gt; 567

&lt;212&gt; DNA

&lt;213&gt; rat

&lt;220&gt;

&lt;223&gt; rat alpha-I partial sequence

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&lt;210&gt; 22

&lt;211&gt; 189

&lt;212&gt; PRT

&lt;213&gt; rat

&lt;220&gt;

&lt;223&gt; rat alpha-I partial sequence

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Gly Val Gln Leu Trp Ala Gly Leu Leu Arg Asn Arg Cys Phe Leu Glu  
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Glu Asn Phe Thr Ile Gln Gly Asp Val Ala Leu Pro Pro Tyr Tyr Gln  
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Pro Glu Glu Asp Asp Glu Met Pro Phe Ile Cys Ser Leu Thr Gly Asp  
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Asn Gly Ile Met Gly Cys His Glu Ile Pro Pro Leu Lys Glu Gln Gly  
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Arg Glu Cys Cys Leu Ser Lys Asp Asp Val Tyr Asp Phe Gly Ala Gly  
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Arg Gln Asp Leu Asn Ala Ser Gly Leu Cys Val Asn Trp Asn Arg Tyr  
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Tyr Asn Val Cys Arg Thr Gly Asn Ala Asn Pro His Lys Gly Ala Ile  
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Asn Phe Asp Asn Ile Gly Tyr Ala Trp Ile Val Ile Phe Gln Val Ile  
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Thr Leu Glu Gly Trp Val Glu Ile Met Tyr Tyr Val Met Asp Ala His  
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Ser Phe Tyr Asn Phe Ile Tyr Phe Ile Leu Leu Ile Ile  
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&lt;211&gt; 7540

&lt;212&gt; DNA

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&lt;400&gt; 23



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Pro Val Ala Ser Arg Ser Ser Thr Thr Cys Pro Gly Pro Gly Ala Ala  
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Gly	Ala	Gly	Ser	Thr	Glu	Lys	Asp	Pro	Gly	Ser	Ala	Asp	Ser	Glu	Ala	65	70	75	80
Glu	Gly	Leu	Pro	Tyr	Pro	Ala	Leu	Ala	Pro	Val	Val	Phe	Phe	Tyr	Leu	85	90	95	
Ser	Gln	Asp	Ser	Arg	Pro	Arg	Ser	Trp	Cys	Leu	Arg	Thr	Val	Cys	Asn	100	105	110	
Pro	Trp	Phe	Glu	Arg	Val	Ser	Met	Leu	Val	Ile	Leu	Leu	Asn	Cys	Val	115	120	125	
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Ala	Val	Glu	Met	Val	Val	Lys	Met	Val	Ala	Leu	Gly	Ile	Phe	Gly	Lys	165	170	175	
Lys	Cys	Tyr	Leu	Gly	Asp	Thr	Trp	Asn	Arg	Leu	Asp	Phe	Phe	Ile	Val	180	185	190	
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&lt;210&gt; 28

&lt;211&gt; 1792

&lt;212&gt; PRT

&lt;213&gt; rat

&lt;400&gt; 28

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```

```

Glu Pro Gly Ile Thr Glu Gln Pro Gly Pro Arg Ser Pro Pro Pro Ser
      20             25             30

```

```

Pro Pro Gly Leu Glu Glu Pro Leu Glu Gly Thr Asn Pro Asp Val Pro
      35             40             45

```

```

His Pro Asp Leu Ala Pro Val Ala Phe Phe Cys Leu Arg Gln Thr Thr
      50             55             60

```

```

Ser Pro Arg Asn Trp Cys Ile Lys Met Val Cys Asn Pro Trp Phe Glu
      65             70             75             80

```

```

Cys Val Ser Met Leu Val Ile Leu Leu Asn Cys Val Thr Leu Gly Met
      85             90             95

```

```

Tyr Gln Pro Cys Asp Asp Met Glu Cys Leu Ser Asp Arg Cys Lys Ile
      100            105            110

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Leu Gln Val Phe Asp Asp Phe Ile Phe Ile Phe Phe Ala Met Glu Met
      115            120            125

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Val Leu Lys Met Val Ala Leu Gly Ile Phe Gly Lys Lys Cys Tyr Leu

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Gly Asp Thr Trp Asn Arg Leu Asp Phe Phe Ile Val Met Ala Gly Met 145 150 155 160		
Val Glu Tyr Ser Leu Asp Leu Gln Asn Ile Asn Leu Ser Ala Ile Arg 165 170 175		
Thr Val Arg Val Leu Arg Pro Leu Lys Ala Ile Asn Arg Val Pro Ser 180 185 190		
Leu Arg Ile Leu Val Asn Leu Leu Leu Asp Thr Leu Pro Met Leu Gly 195 200 205		
Asn Val Leu Leu Leu Cys Phe Phe Val Phe Phe Ile Phe Gly Ile Ile 210 215 220		
Gly Val Gln Leu Trp Ala Gly Leu Leu Arg Asn Arg Cys Phe Leu Glu 225 230 235 240		
Glu Asn Phe Thr Ile Gln Gly Asp Val Ala Leu Pro Pro Tyr Tyr Gln 245 250 255		
Pro Glu Glu Asp Asp Glu Met Pro Phe Ile Cys Ser Leu Thr Gly Asp 260 265 270		
Asn Gly Ile Met Gly Cys His Glu Ile Pro Pro Leu Lys Glu Gln Gly 275 280 285		
Arg Glu Val Cys Leu Ser Lys Asp Asp Val Tyr Asp Phe Gly Ala Gly 290 295 300		
Arg Gln Asp Leu Asn Ala Ser Gly Leu Cys Val Asn Trp Asn Arg Tyr 305 310 315 320		
Tyr Asn Val Cys Arg Thr Gly Asn Ala Asn Pro His Lys Gly Ala Ile 325 330 335		
Asn Phe Asp Asn Ile Gly Tyr Ala Trp Ile Val Ile Phe Gln Val Ile 340 345 350		
Thr Leu Glu Gly Trp Val Glu Ile Met Tyr Tyr Val Met Asp Ala His 355 360 365		
Ser Phe Tyr Asn Phe Ile Leu Leu Ile Ile Val Gly Ser Phe Phe Met 370 375 380		
Ile Asn Leu Cys Leu Val Leu Ile Ala Thr Gln Phe Ser Glu Thr Lys		

385	390	395	400
Gln Arg Asn His Arg Leu Met Leu Glu	Gln Arg Gln Arg Tyr Leu Ser		
405	410	415	
Ser Ser Thr Val Ala Ser Tyr Ala Glu	Pro Gly Asp Cys Tyr Glu Glu		
420	425	430	
Ile Phe Gln Tyr Val Cys His Ile Leu Arg	Lys Ala Lys Arg Arg Ala		
435	440	445	
Leu Gly Leu Tyr Gln Ala Leu Gln Asn Arg	Arg Gln Ala Met Gly Pro		
450	455	460	
Gly Thr Pro Ala Pro Ala Lys Pro Gly	Pro His Ala Lys Glu Pro Ser		
465	470	475	480
His Ser Lys Leu Cys Pro Arg His Ser	Pro Leu Asp Pro Thr Pro His		
485	490	495	
Thr Leu Val Gln Pro Ile Ser Ala Ile	Leu Ala Ser Tyr Pro Ser Ser		
500	505	510	
Cys Pro His Cys Gln His Glu Ala Gly	Arg Arg Pro Ser Gly Leu Gly		
515	520	525	
Ser Thr Asp Ser Gly Gln Glu Gly Ser	Gly Ser Gly Gly Ser Ala Glu		
530	535	540	
Ala Glu Ala Asn Gly Asp Gly Leu Gln	Ser Arg Glu Asp Gly Val Ser		
545	550	555	560
Ser Asp Leu Gly Lys Glu Glu Glu Gln	Glu Asp Gly Ala Ala Arg Leu		
565	570	575	
Cys Gly Asp Val Trp Arg Glu Thr Arg	Lys Lys Leu Arg Gly Ile Val		
580	585	590	
Asp Ser Lys Tyr Phe Asn Arg Gly Ile	Met Met Ala Ile Leu Val Asn		
595	600	605	
Thr Val Ser Met Gly Ile Glu His His	Glu Gln Pro Glu Glu Leu Thr		
610	615	620	
Asn Ile Leu Glu Ile Cys Asn Val Val	Phe Thr Ser Met Phe Ala Leu		
625	630	635	640
Glu Met Ile Leu Lys Leu Ala Ala Phe	Gly Leu Phe Asp Tyr Leu Arg		

645

650

655

Asn Pro Tyr Asn Ile Phe Asp Ser Ile Ile Val Ile Ile Ser Ile Trp  
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 Glu Ile Val Gly Gln Ala Asp Ser Gly Leu Ser Val Leu Arg Thr Ser  
 675 680 685  
 Arg Leu Leu Arg Val Leu Lys Leu Val Arg Phe Met Pro Ala Leu Arg  
 690 695 700  
 Gln Leu Val Val Leu Met Lys Thr Met Asp Asn Val Ala Thr Phe Cys  
 705 710 715 720  
 Met Leu Leu Met Leu Phe Ile Phe Ile Phe Ser Ile Leu Gly Ile Asp  
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 Ile Phe Gly Cys Lys Phe Ser Leu Arg Thr Asp Thr Gly Asp Thr Val  
 740 745 750  
 Pro Asp Arg Lys Asn Phe Asp Ser Leu Leu Trp Ala Ile Val Thr Val  
 755 760 765  
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 Met Ala Ser Thr Thr Pro Trp Ala Ser Leu Tyr Phe Val Ala Leu Met  
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 Thr Phe Gly Asn Tyr Val Leu Phe Asn Leu Leu Val Ala Ile Leu Val  
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 Glu Gly Phe Gln Ala Glu Gly Asp Ala Asn Arg Ser Tyr Ser Asp Glu  
 820 825 830  
 Asp Gln Ser Ser Ser Asn Leu Glu Glu Leu Asp Lys Leu Pro Glu Gly  
 835 840 845  
 Leu Asp Asn Arg Arg Asp Leu Lys Leu Cys Pro Ile Pro Met Thr Pro  
 850 855 860  
 Asn Gly His Leu Asp Pro Ser Leu Pro Leu Gly Ala His Leu Gly Pro  
 865 870 875 880  
 Ala Gly Thr Met Gly Thr Ala Pro Arg Leu Ser Leu Gln Pro Asp Pro  
 885 890 895  
 Val Leu Val Ala Arg Asp Ser Arg Lys Ser Ser Tyr Trp Ser Leu Gly



1155	1160	1165
Ala Ile Phe Val Gly Glu Met Thr Leu Lys Val Val Ser Leu Gly Leu 1170	1175	1180
Tyr Phe Gly Glu Gln Ala Tyr Leu Arg Thr Asp Trp Asn Val Leu Asp 1185	1190	1195 1200
Gly Phe Leu Val Phe Val Ser Ile Ile Asp Ile Val Val Ser Val Ala 1205	1210	1215
Ser Ala Gly Gly Ala Lys Ile Leu Gly Val Leu Arg Leu Leu Arg Thr 1220	1225	1230
Leu Arg Pro Leu Arg Val Ile Ser Arg Ala Pro Gly Leu Lys Leu Val 1235	1240	1245
Val Glu Thr Leu Ile Ser Ser Leu Lys Pro Ile Gly Asn Ile Val Leu 1250	1255	1260
Ile Cys Cys Ala Phe Phe Ile Ile Phe Gly Ile Leu Gly Val Gln Leu 1265	1270	1275 1280
Phe Lys Gly Lys Phe Tyr His Cys Leu Gly Val Asp Thr Arg Asn Ile 1285	1290	1295
Thr Asn Arg Ser Asp Cys Val Ala Ala Asn Tyr Arg Trp Val His His 1300	1305	1310
Lys Tyr Asn Phe Asp Asn Leu Gly Gln Ala Leu Met Ser Leu Phe Val 1315	1320	1325
Leu Ala Ser Lys Asp Gly Trp Val Asn Ile Met Tyr Asn Gly Leu Asp 1330	1335	1340
Ala Val Ala Val Asp Gln Gln Pro Val Thr Asn His Asn Pro Trp Met 1345	1350	1355 1360
Leu Leu Tyr Phe Ile Ser Phe Leu Leu Ile Val Ser Phe Phe Val Leu 1365	1370	1375
Asn Met Phe Val Gly Val Val Val Glu Asn Phe His Lys Cys Arg Gln 1380	1385	1390
His Gln Glu Ala Glu Glu Ala Arg Arg Glu Glu Lys Arg Leu Arg 1395	1400	1405
Arg Leu Glu Lys Lys Arg Arg Tyr Ala Gln Arg Leu Pro Tyr Tyr Ala		



1410	1415	1420
Thr Tyr Cys Pro Thr Arg Leu Leu Ile His Ser Met Cys Thr Ser His 1425                      1430                      1435                      1440		
Tyr Leu Asp Ile Phe Ile Thr Phe Ile Ile Cys Leu Asn Val Val Thr 1445                      1450                      1455		
Met Ser Leu Glu His Tyr Asn Gln Pro Thr Ser Leu Glu Thr Ala Leu 1460                      1465                      1470		
Lys Tyr Cys Asn Tyr Met Phe Thr Thr Val Phe Val Leu Glu Ala Val 1475                      1480                      1485		
Leu Lys Leu Val Ala Phe Gly Leu Arg Arg Phe Phe Lys Asp Arg Trp 1490                      1495                      1500		
Asn Gln Leu Asp Leu Ala Ile Val Leu Leu Ser Val Met Gly Ile Thr 1505                      1510                      1515                      1520		
Leu Glu Glu Ile Glu Ile Asn Ala Ala Leu Pro Ile Asn Pro Thr Ile 1525                      1530                      1535		
Ile Arg Ile Met Arg Val Leu Arg Ile Ala Arg Val Leu Lys Leu Leu 1540                      1545                      1550		
Lys Met Ala Thr Gly Met Arg Ala Leu Leu Asp Thr Val Val Gln Ala 1555                      1560                      1565		
Leu Pro Gln Val Gly Asn Leu Gly Leu Leu Phe Met Leu Leu Phe Phe 1570                      1575                      1580		
Ile Tyr Ala Ala Leu Gly Val Glu Leu Phe Gly Lys Leu Val Cys Asn 1585                      1590                      1595                      1600		
Asp Glu Asn Pro Cys Glu Gly Met Ser Arg His Ala Thr Phe Glu Asn 1605                      1610                      1615		
Ser Ala Arg Ala Phe Leu Thr Leu Phe Gln Val Ser Thr Gly Asp Asn 1620                      1625                      1630		
Trp Asn Gly Ile Met Lys Asp Thr Leu Arg Asp Cys Thr His Asp Glu 1635                      1640                      1645		
Arg Thr Cys Leu Ser Ser Leu Gln Phe Val Ser Pro Leu Tyr Phe Val 1650                      1655                      1660		
Ser Phe Val Leu Thr Ala Gln Phe Val Leu Ile Asn Val Val Val Ala		

1665                      1670                      1675                      1680  
 Val Leu Met Lys His Leu Asp Asp Ser Asn Lys Glu Ala Gln Glu Asp  
                                  1685    1690    1695  
 Ala Glu Met Asp Ala Glu Ile Glu Leu Glu Met Ala His Gly Ser Gly  
                                  1700    1705    1710  
 Pro Cys Pro Gly Pro Cys Pro Gly Pro Cys Pro Cys Pro Cys Pro Cys  
                                  1715    1720    1725  
 Pro Cys Ser Gly Pro Arg Cys Pro Leu Val Thr Trp Gly Ser Gly Ala  
                                  1730    1735    1740  
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                                  1745    1750    1755    1760  
 Arg Thr Ala Ile Arg Cys Trp Thr Pro Arg Val Thr Cys Ala Gly Thr  
                                  1765    1770    1775  
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                                  1780    1785    1790

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 <213> rat

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&lt;400&gt; 30

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&lt;210&gt; 31

&lt;211&gt; 644

&lt;212&gt; PRT

&lt;213&gt; HUMAN

&lt;400&gt; 31

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1

5

10

15

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 Gly Pro Gly Ser Ala Glu Lys Asp Pro Gly Ser Ala Asp Ser Glu Ala  
 35 40 45  
 Glu Gly Leu Pro Tyr Pro Ala Leu Ala Pro Val Val Phe Phe Tyr Leu  
 50 55 60  
 Ser Gln Asp Ser Arg Pro Arg Ser Trp Cys Leu Arg Thr Val Cys Asn  
 65 70 75 80  
 Pro Trp Phe Glu Arg Ile Ser Met Leu Val Ile Leu Leu Asn Cys Val  
 85 90 95  
 Thr Leu Gly Met Phe Arg Pro Cys Glu Asp Ile Ala Cys Asp Ser Gln  
 100 105 110  
 Arg Cys Arg Ile Leu Gln Ala Phe Asp Asp Phe Ile Phe Ala Phe Phe  
 115 120 125  
 Ala Val Glu Met Val Val Lys Met Val Ala Leu Gly Ile Phe Gly Lys  
 130 135 140  
 Lys Cys Tyr Leu Gly Asp Thr Trp Asn Arg Leu Asp Phe Phe Ile Val  
 145 150 155 160  
 Ile Ala Gly Met Leu Glu Tyr Ser Leu Asp Leu Gln Asn Val Ser Phe  
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 Ser Ala Val Arg Thr Val Arg Val Leu Arg Pro Leu Arg Ala Ile Asn  
 180 185 190  
 Arg Val Pro Ser Met Arg Ile Leu Val Thr Leu Leu Leu Asp Thr Leu  
 195 200 205  
 Pro Met Leu Gly Asn Val Leu Leu Leu Cys Phe Phe Val Phe Phe Ile  
 210 215 220  
 Phe Gly Ile Val Gly Val Gln Leu Trp Ala Gly Leu Leu Arg Asn Arg  
 225 230 235 240  
 Cys Phe Leu Pro Glu Asn Phe Ser Leu Pro Leu Ser Val Asp Leu Glu  
 245 250 255  
 Arg Tyr Tyr Gln Thr Glu Asn Glu Asp Glu Ser Pro Phe Ile Cys Ser  
 260 265 270

Gln Pro Arg Glu Asn Gly Met Arg Ser Cys Arg Ser Val Pro Thr Leu  
275 280 285

Arg Gly Asp Gly Gly Gly Gly Pro Pro Cys Gly Leu Asp Tyr Glu Ala  
290 295 300

Tyr Asn Ser Ser Ser Asn Thr Thr Cys Val Asn Trp Asn Gln Tyr Tyr  
305 310 315 320

Thr Asn Cys Ser Ala Gly Glu His Asn Pro Phe Lys Gly Ala Ile Asn  
325 330 335

Phe Asp Asn Ile Gly Tyr Ala Trp Ile Ala Ile Phe Gln Val Ile Thr  
340 345 350

Leu Glu Gly Trp Val Asp Ile Met Tyr Phe Val Met Asp Ala His Ser  
355 360 365

Phe Tyr Asn Phe Ile Tyr Phe Ile Leu Leu Ile Ile Val Gly Ser Phe  
370 375 380

Phe Met Ile Asn Leu Cys Leu Val Val Ile Ala Thr Gln Phe Ser Glu  
385 390 395 400

Thr Lys Gln Arg Glu Ser Gln Leu Met Arg Glu Gln Arg Val Arg Phe  
405 410 415

Leu Ser Asn Ala Ser Thr Leu Ala Ser Phe Ser Glu Pro Gly Ser Cys  
420 425 430

Tyr Glu Glu Leu Leu Lys Tyr Leu Val Tyr Ile Leu Arg Lys Ala Ala  
435 440 445

Arg Arg Leu Ala Gln Val Ser Arg Ala Ala Gly Val Arg Val Gly Leu  
450 455 460

Leu Ser Ser Pro Ala Pro Leu Gly Gly Gln Glu Thr Gln Pro Ser Ser  
465 470 475 480

Ser Cys Ser Arg Ser His Arg Arg Leu Ser Val His His Leu Val His  
485 490 495

His His His His His His His Tyr His Leu Gly Asn Gly Thr Leu  
500 505 510

Arg Ala Pro Arg Ala Ser Pro Glu Ile Gln Asp Arg Asp Ala Asn Gly  
515 520 525

Ser Arg Arg Leu Met Leu Pro Pro Pro Ser Thr Pro Ala Leu Ser Gly  
530 535 540

Ala Pro Pro Gly Gly Ala Glu Ser Val His Ser Phe Tyr His Ala Asp  
545 550 555 560

Cys His Leu Glu Pro Val Arg Cys Gln Ala Pro Pro Pro Arg Ser Pro  
565 570 575

Ser Glu Ala Ser Gly Arg Thr Val Gly Ser Gly Lys Val Tyr Pro Thr  
580 585 590

Val His Thr Ser Pro Pro Pro Glu Thr Leu Lys Glu Lys Ala Leu Val  
595 600 605

Glu Val Ala Ala Ser Ser Gly Pro Pro Thr Leu Thr Ser Leu Asn Ile  
610 615 620

Pro Pro Gly Pro Tyr Ser Ser Met His Lys Leu Leu Glu Thr Gln Ser  
625 630 635 640

Thr Gly Ala Cys

<210> 32

<211> 1608

<212> DNA

<213> HUMAN

<400> 32

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cggcgttggt gggggcgctc ccggagagcc ccggggcgcc gggacgcgag gcggagcggg 180  
ggtccgagct cggcggtgtc ccctccgaga gcccgggcgc cgagcgcggc gcggagctgg 240  
gtgccgacga ggagcagcgc gtcccgtacc cggccttggc ggccacggtc ttcttctgcc 300  
tcggtcagac cagcgggcgc cgcagctggt gcctccggct ggtctgcaac ccatgggttcg 360  
agcacgtgag catgctggta atcatgctca actgctgac cctgggcatg ttccggccct 420  
gtgaggacgt tgagtgcggc tccgagcgt gcaacatcct ggaggccttt gacgccttca 480  
ttttcgccct ttttgcgggt gagatggtca tcaagatggt ggccctgggg ctgttcgggc 540  
agaagtgtta cctgggtgac acgtggaaca ggctggattt cttcatcgtc gtggcgggca 600  
tgatggagta ctggttgac ggacacaacg tgagcctctc ggctatcagg accgtgcggg 660  
tgctgcggcc cctccgcgcc atcaaccgcg tgcctagcat gcggatcctg gtcactctgc 720  
tgctggatac gctgcccacg ctccgggaacg tccttctgct gtgcttcttc gtcttcttca 780  
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acagtgcctt tgtcaggaac aacaacctga ccttctctgc gccgtactac cagacggagg 900  
agggcgagga gaaccgctt atctgtcct cagcccgaga caacggcatg cagaagtgtc 960  
cgcacatccc cggccgcccgc gagctgcgca tgccctgcac cctgggctgg gaggcctaca 1020

cgcagccgca ggccgagggg gtggggcgctg cgcgcaacgc ctgcatcaac tgggaaccagt 1080  
 actacaacgt gtgccgctcg ggtgactcca acccccacaa cggtgccatc aacttcgaca 1140  
 acatcggtta cgcctggatc gccatcttcc aggtgatcac gctggaaggc tgggtggaca 1200  
 tcatgtacta cgtcatggac gccactcat tctacaactt catctatttc atcctgctca 1260  
 tcatcgtggg ctcttcttc atgatcaacc tgtgcctggt ggtgattgcc acgcagttct 1320  
 cggagacgaa gcagcgggag agtcagctga tgcgggagca gcgggcacgc cacctgtcca 1380  
 acgacagcac gctggccagc ttctccgagc ctggcagctg ctacgaagag ctgctgaagt 1440  
 acgtgggcca catattccgc aaggtcaagc ggcgcagctt gcgcctctac gcccgctggc 1500  
 agagccgctg gcgcaagaag gtggacccca gtgctgtgca aggccagggt cccgggcacc 1560  
 gccagcgccg ggcaggcagg cacacagcct cgggtgcacca cctggtct 1608

&lt;210&gt; 33

&lt;211&gt; 518

&lt;212&gt; PRT

&lt;213&gt; HUMAN

&lt;400&gt; 33

Met Thr Glu Gly Ala Arg Ala Ala Asp Glu Val Arg Val Pro Leu Gly  
 1 5 10 15  
 Ala Pro Pro Pro Gly Pro Ala Ala Leu Val Gly Ala Ser Pro Glu Ser  
 20 25 30  
 Pro Gly Ala Pro Gly Arg Glu Ala Glu Arg Gly Ser Glu Leu Gly Val  
 35 40 45  
 Ser Pro Ser Glu Ser Pro Ala Ala Glu Arg Gly Ala Glu Leu Gly Ala  
 50 55 60  
 Asp Glu Glu Gln Arg Val Pro Tyr Pro Ala Leu Ala Ala Thr Val Phe  
 65 70 75 80  
 Phe Cys Leu Gly Gln Thr Thr Arg Pro Arg Ser Trp Cys Leu Arg Leu  
 85 90 95  
 Val Cys Asn Pro Trp Phe Glu His Val Ser Met Leu Val Ile Met Leu  
 100 105 110  
 Asn Cys Val Thr Leu Gly Met Phe Arg Pro Cys Glu Asp Val Glu Cys  
 115 120 125  
 Gly Ser Glu Arg Cys Asn Ile Leu Glu Ala Phe Asp Ala Phe Ile Phe  
 130 135 140  
 Ala Phe Phe Ala Val Glu Met Val Ile Lys Met Val Ala Leu Gly Leu  
 145 150 155 160  
 Phe Gly Gln Lys Cys Tyr Leu Gly Asp Thr Trp Asn Arg Leu Asp Phe

165

170

175

Phe Ile Val Val Ala Gly Met Met Glu Tyr Ser Leu Asp Gly His Asn  
 180 185 190  
 Val Ser Leu Ser Ala Ile Arg Thr Val Arg Val Leu Arg Pro Leu Arg  
 195 200 205  
 Ala Ile Asn Arg Val Pro Ser Met Arg Ile Leu Val Thr Leu Leu Leu  
 210 215 220  
 Asp Thr Leu Pro Met Leu Gly Asn Val Leu Leu Leu Cys Phe Phe Val  
 225 230 235 240  
 Phe Phe Ile Phe Gly Ile Val Gly Val Gln Leu Trp Ala Gly Leu Leu  
 245 250 255  
 Arg Asn Arg Cys Phe Leu Asp Ser Ala Phe Val Arg Asn Asn Asn Leu  
 260 265 270  
 Thr Phe Leu Arg Pro Tyr Tyr Gln Thr Glu Glu Gly Glu Glu Asn Pro  
 275 280 285  
 Phe Ile Cys Ser Ser Arg Arg Asp Asn Gly Met Gln Lys Cys Ser His  
 290 295 300  
 Ile Pro Gly Arg Arg Glu Leu Arg Met Pro Cys Thr Leu Gly Trp Glu  
 305 310 315 320  
 Ala Tyr Thr Gln Pro Gln Ala Glu Gly Val Gly Ala Ala Arg Asn Ala  
 325 330 335  
 Cys Ile Asn Trp Asn Gln Tyr Tyr Asn Val Cys Arg Ser Gly Asp Ser  
 340 345 350  
 Asn Pro His Asn Gly Ala Ile Asn Phe Asp Asn Ile Gly Tyr Ala Trp  
 355 360 365  
 Ile Ala Ile Phe Gln Val Ile Thr Leu Glu Gly Trp Val Asp Ile Met  
 370 375 380  
 Tyr Tyr Val Met Asp Ala His Ser Phe Tyr Asn Phe Ile Tyr Phe Ile  
 385 390 395 400  
 Leu Leu Ile Ile Val Gly Ser Phe Phe Met Ile Asn Leu Cys Leu Val  
 405 410 415  
 Val Ile Ala Thr Gln Phe Ser Glu Thr Lys Gln Arg Glu Ser Gln Leu



420

425

430

Met Arg Glu Gln Arg Ala Arg His Leu Ser Asn Asp Ser Thr Leu Ala  
 435 440 445

Ser Phe Ser Glu Pro Gly Ser Cys Tyr Glu Glu Leu Leu Lys Tyr Val  
 450 455 460

Gly His Ile Phe Arg Lys Val Lys Arg Arg Ser Leu Arg Leu Tyr Ala  
 465 470 475 480

Arg Trp Gln Ser Arg Trp Arg Lys Lys Val Asp Pro Ser Ala Val Gln  
 485 490 495

Gly Gln Gly Pro Gly His Arg Gln Arg Arg Ala Gly Arg His Thr Ala  
 500 505 510

Ser Val His His Leu Val  
 515

&lt;210&gt; 34

&lt;211&gt; 1080

&lt;212&gt; DNA

&lt;213&gt; HUMAN

&lt;400&gt; 34

gcagtgtcat gtctctaggg aggatgagct atgaccagcg ctccctgtcc agtccccgga 60  
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 acagcctcaa gcacaagccg ccgtcggcgg agcatgagtc cctgctctct gcggagcggc 180  
 gcggcggcgc ccgggtctgc gaggttgccg cggacgaggg gccgcgcgg gccgcacccc 240  
 tgcacacccc acacgcccac cacattcatc acggggccca tctggcgcac cgccaccgcc 300  
 accaccgccg gacgtgttcc ctcgacaaca gggactcggg ggacctggcc gagctggtgc 360  
 ccgcgggtggg cgcccacccc cgggcccgcct ggagggcggc agggccggcc cccgggcatg 420  
 aggactgcaa tggcaggatg cccagcatcg ccaaagacgt cttcaccaag atgggcgacc 480  
 gcggggatcg cggggaggat gaggaggaaa tcgactacac cctgtgcttc cgcgtccgca 540  
 agatgatcga cgtctataag cccgactggt gcgaggtccg cgaagactgg tctgtctacc 600  
 tcttctctcc cgagaacagg ttccgggtcc tgtgtcagac cattattgcc cacaaactct 660  
 tcgactacgt cgctctggcc ttcatctttc tcaactgcat caccatcgcc ctggagcggc 720  
 ctcagatcga ggccggcagc accgaacgca tctttctcac cgtgtccaac tacatcttca 780  
 cggccatctt cgtgggcgag atgacattga aggtagtctc gctgggcctg tacttcggcg 840  
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 tcatcgacat cgtggtgtcc ctggcctcag ccgggggagc caagatcttg ggggtcctcc 960  
 gagtcttgcg gctcctgcgc accctacgcc ccctgcgtgt catcagccgg gcgccggggc 1020  
 tgaagctggt ggtggagaca ctcctctcct ccdtcaagcc catcggaac atcgtgctca 1080

&lt;210&gt; 35

&lt;211&gt; 359

<212> PRT  
<213> HUMAN

<400> 35

Ser Val Met Ser Leu Gly Arg Met Ser Tyr Asp Gln Arg Ser Leu Ser  
1 5 10 15

Ser Ser Arg Ser Ser Tyr Tyr Gly Pro Trp Gly Arg Ser Ala Ala Trp  
20 25 30

Ala Ser Arg Arg Ser Ser Trp Asn Ser Leu Lys His Lys Pro Pro Ser  
35 40 45

Ala Glu His Glu Ser Leu Leu Ser Ala Glu Arg Gly Gly Gly Ala Arg  
50 55 60

Val Cys Glu Val Ala Ala Asp Glu Gly Pro Pro Arg Ala Ala Pro Leu  
65 70 75 80

His Thr Pro His Ala His His Ile His His Gly Pro His Leu Ala His  
85 90 95

Arg His Arg His His Arg Arg Thr Leu Ser Leu Asp Asn Arg Asp Ser  
100 105 110

Val Asp Leu Ala Glu Leu Val Pro Ala Val Gly Ala His Pro Arg Ala  
115 120 125

Ala Trp Arg Ala Ala Gly Pro Ala Pro Gly His Glu Asp Cys Asn Gly  
130 135 140

Arg Met Pro Ser Ile Ala Lys Asp Val Phe Thr Lys Met Gly Asp Arg  
145 150 155 160

Gly Asp Arg Gly Glu Asp Glu Glu Glu Ile Asp Tyr Thr Leu Cys Phe  
165 170 175

Arg Val Arg Lys Met Ile Asp Val Tyr Lys Pro Asp Trp Cys Glu Val  
180 185 190

Arg Glu Asp Trp Ser Val Tyr Leu Phe Ser Pro Glu Asn Arg Phe Arg  
195 200 205

Val Leu Cys Gln Thr Ile Ile Ala His Lys Leu Phe Asp Tyr Val Val  
210 215 220

Leu Ala Phe Ile Phe Leu Asn Cys Ile Thr Ile Ala Leu Glu Arg Pro  
225 230 235 240

Gln Ile Glu Ala Gly Ser Thr Glu Arg Ile Phe Leu Thr Val Ser Asn  
245 250 255

Tyr Ile Phe Thr Ala Ile Phe Val Gly Glu Met Thr Leu Lys Val Val  
260 265 270

Ser Leu Gly Leu Tyr Phe Gly Glu Gln Ala Tyr Leu Arg Ser Ser Trp  
275 280 285

Asn Val Leu Asp Gly Phe Leu Val Phe Val Ser Ile Ile Asp Ile Val  
290 295 300

Val Ser Leu Ala Ser Ala Gly Gly Ala Lys Ile Leu Gly Val Leu Arg  
305 310 315 320

Val Leu Arg Leu Leu Arg Thr Leu Arg Pro Leu Arg Val Ile Ser Arg  
325 330 335

Ala Pro Gly Leu Lys Leu Val Val Glu Thr Leu Ile Ser Ser Leu Lys  
340 345 350

Pro Ile Gly Asn Ile Val Leu  
355